© 2010 Revista Nefrología. Official Publication of the Spanish Nephrology Society

# Growth factors and renal regeneration

M. Flaquer<sup>1</sup>, P. Romagnani<sup>2</sup>, J.M. Cruzado<sup>3</sup>

<sup>1</sup> Experimental Nephrology Laboratory. Hospital de Bellvitge. IDIBELL. L'Hospitalet de Llobregat, Barcelona (Spain)

<sup>2</sup> Excellence Centre for Research, Transfer and High Education for the Development of DE NOVO Therapies (DENOTHE). University of Florence. Florence (Italy)

<sup>3</sup> Nephrology Department. Bellvitge Hospital. IDIBELL. L'Hospitalet de Llobregat, Barcelona (Spain)

#### Nefrologia 2010;30(4):385-93

doi:10.3265/Nefrologia.pre2010.Jun.10463

#### ABSTRACT

Cell replenishment is critical for adult tissue repair after damage. In some organs this process is facilitated by stem cells. In contrast to the liver, the kidney has limited regeneration capacity and has even been considered over several years as not being able to regenerate itself. Nevertheless, there are several recent studies suggesting the presence of stem cells in the adult kidney. Stem cell renal niches have been identified in the renal papillae in animals as well as in the urinary pole of the Bowman's capsule in humans (CD24+CD133+ stem cells). Although these cells may contribute to organ regeneration, how these cells exert this effect and their role after kidney injury is not known. Nevertheless, renal stem cells may be therapeutic targets for treatment of renal diseases. On the other hand, bone-marrow-derived stem cells may also contribute to renal repair, particularly mesenchymal stem cells. However, the mechanism for producing such effect has not been elucidated. Some studies suggest there is cell fusion between bone marrow and resident tubular cells: others suggest that bone marrow cells are able to differentiate in resident cells, while some authors propose bone marrow cells facilitate organ regeneration by a paracrine action; that is by secreting growth factors such as HGF, VEGF and IGF1. All these secreted molecules would provide a regenerative milieu able to constrain renal damage and to amplify stem cells migration to the damaged organ.

**Key words:** Renal regeneration, Stem cells, Hepatocyte growth factor, Bone marrow.

**Correspondence:** José María Cruzado Garrit Servicio de Nefrología. Hospital de Bellvitge. IDIBELL. Feixa Llarga, s/n. 08907 L'Hospitalet de Llobregat, Barcelona. Spain. Tel: +34 93 2607602 jmcruzado@bellvitgehospital.cat

#### Factores de crecimiento y regeneración renal

#### RESUMEN

Cuando se produce un daño en un tejido adulto, el proceso de renovación celular continuada es crítico y crucial para la reparación del mismo y, en determinados órganos, se facilita por la presencia de células madre o progenitoras. El riñón, a diferencia de otros órganos como el hígado, es de regeneración lenta. Incluso ha sido considerado durante años como incapaz de regenerarse. Sin embargo, varios estudios han demostrado que existen posibles nichos de células madre renales en la papila renal, progenitores tubulares o progenitores renales CD24+CD133+ localizados en el polo urinario de la cápsula de Bowman. Estas células podrían participar teóricamente en la reparación de la lesión renal. Sin embargo, todavía no se ha demostrado de forma precisa cuál sería su papel ni cómo actuarían después del daño. Aún así, estas células madre renales podrían ser dianas terapéuticas para el remodelado del tejido renal dañado. Por otro lado, se ha postulado que las células madre derivadas de la médula ósea podrían participar en la regeneración renal, especialmente las de estirpe mesenguimal. Sin embargo, tampoco se conoce con exactitud el modo en que actuarían. Hay estudios que sugieren la existencia de fusión celular entre estas células y células residentes, otros apuntan a su diferenciación en células renales, mientras que otros sugieren una acción paracrina responsable del efecto reparador a través de la secreción de factores de crecimiento como HGF, VEGF y IGF-1. Todas estas moléculas secretadas proporcionarían un entorno regenerativo que limitaría el área del daño y que facilitaría la migración de las células madre.

**Palabras clave:** Regeneración renal, Células madre, Factor de crecimiento de los hepatocitos, Médula ósea.

#### INTRODUCTION

The incidence and prevalence of chronic kidney disease (CKD) continue to rise in such a way that it is considered a worldwide public health threat.<sup>12</sup> Many patients with chronic

nephropathy go on to develop terminal chronic kidney failure (TCKF), with diabetes being the most common cause both in Spain and in most western countries. The progression of chronic kidney disease towards loss of function and renal sclerosis also appears in many glomerular, interstitial and vascular chronic nephropathies. Essentially, it is thought that, once enough renal mass is lost, the residual nephrons suffer from intraglomerular hypertension, a phenomenon which produces local activation of, among others, the reninangiotensin-aldosterone system (RAAS), inducing TGF-beta 1 and the production of extracellular matrix which finally accelerates the loss of renal mass. This physiopathological concept, known as the hyperfiltration theory,<sup>3</sup> is the base of clinical testing of the usefulness of the blockade of RAAS, both in diabetic nephropathy and other chronic nephropathies. However, despite the significant advances that these treatments have brought about (they are able to reduce or stabilise the rate of the loss of renal function), the number of incident patients requiring kidney replacement treatment, whether peritoneal dialysis, haemodialysis or kidney transplants, continues to rise. Within this context of a shortage of treatments or strategies able to induce a decline in chronic nephropathy, the study of renal regeneration mechanisms acquires a great deal of interest.4,5

#### **KIDNEY REGENERATION**

When adult tissue is damaged, the continuous cell renewal process is crucial for its maintenance and, in certain organs, this is achieved by the presence of stem cells. Stem cells enable periodic cell renewal or regeneration when tissue is damaged, and they have the capacity for cell renewal through mitotic divisions or for differentiation into the cell lines of the corresponding organ. Furthermore, some adult stem cells from bone marrow are able to differentiate into more than one type of cell (mesenchymal and hematopoietic). In general, adult stem cells are a self-renewing cell population: they are quiescent cells that divide asymmetrically during tissue regeneration, on the one hand, into stem cells and, on the other, into transit-amplifying cells which proliferate, differentiate and lastly, reconstruct damaged tissue.6 One of the ways to identify stem cells in solid organs is to stain them with bromodeoxyuridine (BrdU). Quiescent cells, which do not divide, maintain the high levels of BrdU deposited in their genome, whereas the dividing, more mature stem cells steadily dilute the BrdU incorporated into their genomes as they proliferate.

The kidneys are traditionally considered organs which are unable to regenerate. Yet, they possess a certain degree of regeneration which varies according to the species. Some cartilaginous fish form nephrons during adulthood, while mammals have lost this capacity. In

fact, humans do not form new nephrons after 36 weeks of gestation.7 The kidneys are one of the few organs to undergo mesenchymal-epithelial transition (MET) during their development.8 This process is controlled by growth factors such as hepatocyte growth factor (HGF) and bone morphogenetic protein-7 (BMP-7), amongst many others. Therefore, kidney development in mammals requires a conversion process of the metanephric mesenchymal cells into polarised epithelial cells.9 As mentioned above, when sufficiently extensive chronic renal injury occurs, whatever the cause, kidney function worsens inexorably until reaching TCKF, with no treatment available to reverse this process.10 One of the processes taking place in the progression of nephropathies is epithelial-mesenchymal transition capable of producing extracellular matrix. To be precise, this is the inverse process to that taking place during foetal development of the kidney.

Renal regeneration could be approached using different strategies, such as the administration of growth factors capable of reversing epithelial-mesenchyma transition, and even by the mobilisation or infusion of endogenous (from the kidney itself) or exogenous (from bone marrow) stem cells. However, this is an extremely difficult challenge. Kidneys have a very complex architecture, great cellular heterogeneity and their cell renewal is slow. They have over 24 types of mature stem cell distributed in vascular, interstitial, glomerular and tubular compartments.8 All of this complicates the search for adult stem cells<sup>11</sup> capable of repairing the kidney by replacing damaged cells. In any case, renal regeneration requires very precise mechanisms capable of directing the repair of each of the damaged renal compartments. Studies have been performed supporting the presence of stem cells in adult kidneys, showing that these cells have an intrinsic function. However, the participation of stem cells derived from bone marrow is not so clear (Table 1).<sup>12-17</sup>

#### STEM CELLS AND RENAL REGENERATION

#### **Renal stem cells**

Kidneys have a very complex structure and a very low degree of regeneration compared with other organs. This makes it difficult to study the existence of renal stem cell niches and to investigate how they participate in organ repair. Overcoming these difficulties with different strategies, some stem cell niches have been proposed in different renal compartments. Maeshima et al.<sup>12</sup> described a population of tubular progenitors with regenerative properties which proliferate and differentiate in epithelial cells during tubular regeneration. In fact, the origin of the cells that replace the damaged tubular epithelial cells

Table 1. Participation of endogenous (renal) and exogenous (derived from bone marrow) stem cells in the regeneration of kidney tissue

Damaged Cell	Stem Cell	Repair mechanism	Author and reference
Tubular cells	Renal	Proliferation and differentiation in epithelial cells	Maeshima, et al. <sup>12</sup>
			Duffield, et al. <sup>13</sup>
			Lin, et al. <sup>14</sup>
			Humpphreys, et al.17
Papilla cells	Renal	Cellular niche	Oliver, et al.15
Bowman's capsule cells	Renal	Cellular niche	Sagrinati, et al. <sup>16</sup>
Tubular cells	BMDC	Cellular fusion	Held, et al. <sup>33</sup>
Mesangial cells	BMDC	Cellular differentiation	Imasawa, et al. <sup>71</sup>
Mesangial cells	BMDC (HSC)	Formation of mesangial cells	Masuya, et al.72
Glomerular endothelial cells	BMDC	Glomerular integration and microvascular repair	Rookmaaker, et al.73

Several authors describe the different types of renal cell that participate in kidney regeneration. They also mention the existence of a niche of stem cells in the renal papillae and in the Bowman's capsule. Besides, other studies show that BMDC play a role in mesangial, renal endothelial and tubular mantenaince and repair, by means of cell fusion and differentiation.

is not known, but some studies suggest that they are from the kidneys and not from bone marrow.<sup>13,14</sup> Oliver et al<sup>15</sup> identified the renal papillae as a stem cell niche in the adult kidney. They observed a group of cells in this area that retained BrdU. After ischemic damage, they found that these cells entered a cell cycle, and thus the BrdU stains disappeared. Furthermore, these cells were able to form spheres in vitro. However, these cells being located in the renal papillae creates doubts about how they are able to repopulate the most proximal segments of the nephron. Recent studies in humans have identified a subset of renal progenitors CD24+CD133+ in the Bowman's capsule,16 near to the tubular pole. This location would allow them to repair tubular and glomerular epithelial cells. It has been described that the progenitor cells CD24+CD133+ have the capacity to differentiate, providing a regenerative mechanism for damaged epithelial cells in the kidneys.<sup>18,19</sup> The existence of these kidney epithelial progenitor cells provides a possible explanation for the regression of kidney lesions. The damage repair process probably requires the capacity to slow the fibrotic response, so progenitor cells should be able to regenerate tissue and, at the same time, prevent extracellular matrix accumulation<sup>18</sup> by means of their capacity to secrete growth factors, as will be detailed below. In another study, Appel et al<sup>20</sup> postulated that, as podocytes have no self-renewal capacity, the glomerular parietal epithelial cells (which proliferate and are adjacent to the podocytes) might migrate to the glomerular capillary and differentiate into podocytes. Although several niches of renal stem cells have been identified, it is still not known what their role is and how they behave in repairs after a kidney injury. Still, renal stem cells could be therapeutic targets for remodelling damaged kidney tissue.21

#### Bone marrow stem cells

Bone marrow contains different types of stem cells, including haematopoietic (HSCs) and mesenchymal (MSCs) stem cells and endothelial progenitor cells. HSCs express surface markers such as Sca-1, c-kit, CD90 in mice, and CD34, CD133, CXCR4 and CD150 in humans, and can differentiate into any type of adult blood cell. Besides creating a support environment for HSCs, MSCs are able to differentiate into various types of mesenchymal cells such as bone, cartilage, muscle, neurons, hepatocytes and adipose tissue.<sup>22-25</sup> They can adhere to plastic and express surface markers such as CD90, CD73, CD105, CD44 and CD29. MSCs also express growth factors such as VEGF, HGF and IGF-1, as well as antiapoptotic cytokines. At present the role of marrow bone stem cells in the kidney regeneration after damage is being investigated. Cell therapy is one of the fields of greatest interest in biomedicine at this time, so much so that the use of these multipotent cells to re-establish damage organ function has generated remarkable expectation.

The most common technique for studying the plasticity of bone marrow cells is bone marrow transplantation (BMT). The receptor's bone marrow cells are replaced by those of the donor and, once chimerism is established, the donor cells can be identified by different strategies. Among these we can highlight the identification of the Y chromosome in a female receptor, the expression of molecules such as beta-galactosidase, luciferase or enhanced green fluorescent protein (EGFP), or a function being reestablished in knockout animal model.<sup>26</sup> To check the cell type (tubular, mesangial, etc.) that the marrow bone cells have given rise to, the use of specific protein staining with immunohistochemistry, immunofluorescence and analysis with a confocal microscope are common.

A significant number of glomerulopathies begin with podocyte injury or loss. Podocytes are cells with complex interdigitations which participate in the synthesis of components of the glomerular basement membrane (GBM), collagen IV being one of the most important. Several studies have suggested the integration of cells derived from bone marrow as functional podocytes. Studies have been performed with mice models of Alport syndrome. The mice had mutations in the gene which codifies for the alpha chain of the collagen IV, bringing about defects in the GBM, proteinuria and kidney failure. Prodromidi et al.27 and Sugimoto et al.28 observed that bone marrow cells contribute to the regeneration of podocytes in damaged glomeruli, leading to the expression of the collagen IV alpha-3 chain being reestablished and a decrease in proteinuria. In a study with mice published a few years ago, the BMT from obese diabetic db/db mice in healthy non-diabetic mice transferred diabetic nephropathy to the receptor mice without them becoming hyperglycaemic. The authors postulated that the glomerulus was probably repopulated by mesangial and endothelial cells from the db/db donor mice and that these were responsible for the albuminuria and glomerulosclerosis that the receptor mice developed.<sup>29</sup>

On the other hand, there are authors who suggest that bone marrow cells participate in renal regeneration by fusing with the renal cells themselves. In fact, studies in livers have shown that hepatocytes generated after liver damage are formed by cellular fusion, and not by differentiation of hematopoietic stem cells.<sup>30-32</sup> Therefore, a possible fusion between bone marrow stem cells and epithelial tubular cells is postulated. Held et al.<sup>33</sup> observed that, after injury, tubular epithelial cells are generated by the fusion of hematopoietic cells and existing proximal tubular cells, and not by transdifferentiation. However, this is still a very controversial matter since several studies suggest a paracrine/endocrine action of endogenous stem cells instead of direct repopulation of the damaged nephrons.<sup>34</sup> In short, besides the possible role of endogenous stem cells (renal), other studies support the idea of differentiation and/or fusion of cells from bone marrow into precursor cells of damaged kidney cells (figure 1).

#### **GROWTH FACTORS**

Tubular epithelial cells which survive damage secrete growth factors which could interact with resident cells and renal and extrarenal stem cells, accelerating tubular repair mechanisms. The tubular epithelium exists in a relatively quiescent to slowly replicating state (also a characteristic of stem cells), but, on the other hand, it has a remarkable morphogenic regeneration capacity after severe toxic or ischemic aggression.<sup>35</sup> Although some studies show how stem cells migrate to damaged tissue,<sup>36,37</sup> most authors do not support the idea of the integration of these cells into injured organs. In this respect, Duffield et al.<sup>13,38</sup> showed that kidney repair is independent of the participation of cells derived from bone marrow, something also observed by Lin et al.<sup>14</sup> On the other hand, Morigi et al<sup>39</sup> showed how the infusion of human mesenchymal cells from bone marrow led to a decrease in proximal tubular damage and improved kidney function in mice. Several studies have been performed which have not been able to verify the differentiation of stem cells into epithelial cells, but others have described how stem cells contribute to renal recovery. Thus, it was proposed that cell migration only facilitates regeneration because of an endocrine/paracrine effect<sup>40,41</sup> and it is the kidney cells themselves that reestablish the tubular epithelium.<sup>13,14,38,42</sup>

# Interaction between stem cells, resident kidney cells and growth factors

The interaction between mesenchymal and epithelial cells and growth factors is fundamental for nephrogenesis and for maintaining the integrity of adult organs.<sup>43,44</sup> This reciprocated interaction between mesenchymal-epithelial cells is a key factor in renal regeneration after damage.

Studies using animal models of acute kidney injury have been performed administering growth factors such as epidermal growth factor (EGF), hepatocyte growth factor (HGF) or insulin-like growth factor 1 (IGF-1). They observed a decrease in mortality due to kidney function being restored and normalised.<sup>45</sup> In fact, it is well known that tubular epithelial cells that survive damage secrete growth factors and cytokines involved in kidney repair mechanisms. On the other hand, it seems to have been proven that MSC have a protective effect, especially in acute kidney injury models, thanks to their ability to express growth factors such as VEGF, HGF and IGF-1, which facilitate recovery from kidney injury.46,47 The mechanism of action of this system could have several modes: autocrine (the kidney cells themselves secrete growth factors), paracrine (renal and bone marrow stem cells) and endocrine (soluble circulating factors). A brief review of some of these factors is given below.

Glial cell line-derived neurotrophic growth factor (GDNF). This factor is involved in renal organogenesis. The exogenous administration of GDNF protects against ischemic kidney injury in a mouse and accelerates repair mechanisms. *In vitro*, GDNF induces MSC migration and inhibits MSC apoptosis.<sup>48</sup>

**Epidermal growth factor (EGF).** This factor is synthesised in the renal epithelium and increases after



Figure 1. Stem cells and renal renegeneration.

kidney damage.<sup>49</sup> It exerts different actions on several types of cells, such as migration and proliferation.<sup>50,51</sup> EGF has been shown to induce cellular proliferation and MSC migration *in vitro*.<sup>52</sup>

Hepatocyte growth factor (HGF). It is a heterodimer consisting of 60-kDa alpha-chain and a 34-kDa betachain. The interaction of HGF and its c-Met receptor leads to the activation of the tyrosine kinase pathway, giving rise to mitogenic and angiogenic activity in several types of cells, particularly in epithelial and endothelial cells.55 Furthermore, HGF has anti-apoptotic and antifibrotic effects. The anti-apoptotic effect is directly related to the phosphatidylinositol-3 kinase-Akt signal pathway,56 while the antifibrotic effect is linked to its antagonist action on TGF beta-1.57 HGF modulates the balance between extracellular matrix synthesis and degradation, increasing expression from matrix metalloproteases (MMP) and reducing production of MMP inhibitors (TIMP). Furthermore, HGF suppresses the effect of TGF beta-1 by blocking the TGF beta/Smad

Nefrologia 2010;30(4):385-93

pathway.58 HGF is able to counteract the profibrotic action of TGF beta-1 in different renal cells through different mechanisms of action, amongst which the inhibition of epithelium-mesenchyma transition stands out. It is also known that TGF beta-1 and HGF inhibit each other's synthesis in a reciprocal fashion<sup>59</sup> and that HGF also down-regulates the expression of TGF beta-1 receptors in vivo. Some authors described a decrease in TGF beta-1 using an exogenous HGF supplement in several chronic damage models.<sup>60-62</sup> It is worth pointing out that HGF also has an effect on bone marrow cells, attracting stem cells to the site of the damage. It is not known if HGF has a cellular mobilisation and/or localisation effect on these cells. In a study performed using a model of liver damage, Kollet et al<sup>64</sup> showed the effect of HGF on the recruitment of hematopoietic cells in liver damage. When the liver is damaged (by irradiation or inflammation) there is an increase in the expression of SDF-1 and MMP-9 activity, giving rise to the recruitment of hematopoietic progenitor cells mediated by SDF-1. In another study performed using a mouse model of CCl<sub>4</sub>-induced hepatic fibrosis,<sup>65</sup> it was

observed that HGF per se did not increase the expression of MMP-9. Treatment with G-CSF is used to promote the recruitment of cells from bone marrow. Overexpression of HGF together with treatment with G-CSF synergistically increased MMP-9 in fibrotic liver while increasing the number of cells from bone marrow and liver cells expressing MMP-9. In addition, it is known that the inhibition of HGF activity leads to impaired tissue repair.<sup>57,66</sup>

**Vascular endothelial growth factor (VEGF).** This is a factor which regulates vascular growth both in normal and damaged tissue. Mesenchymal cells can secrete this factor.<sup>67,69</sup> Renal

ischemia inhibits the expression of VEGF through diverse mechanisms, shifting the balance from a proangiogenic to an antiangiogenic environment, thus inhibiting kidney repair. MSCs express VEGF and could exert a renoprotective, paracrine action which facilitates recovery from acute kidney injury. It has even been postulated that giving high doses of erythropoietin in a model of endothelial lesion attenuates the damage by liberating VEGF.<sup>70</sup>

In short, certain growth factors, many of which are involved in renal embryogenesis, are able to directly induce a certain degree of tissue repair, while possibly acting on resident stem cells, facilitating their



Figure 2. Growth factors and renal renegeneration.

differentiation and even contributing to the recruitment in the kidneys of stem cells from bone marrow which, directly or through the secretion of growth factors, play a part in renal regeneration (figure 2).

#### Acknowledgements

This research has been possible thanks to the ISCIII/FIS P106/0582 and PS09/01630 grants. Maria Flaquer has received an IDIBELL predoctoral grant.

### **KEY CONCEPTS**

- The incidence of CKD continues to increase and many patients go on to develop TCKF despite the advances in renoprotection. Because of this, there is a huge interest in the study of renal regeneration mechanisms, looking for future clinical applications.
- 2. Possible renal stem cell niches have been described in the renal papillae and the urinary pole of the Bowman's capsule, and tubular progenitor cells have also been observed. It has not been proven irrefutably that stem cells from bone marrow differentiate and integrate in vivo like adult renal cells.
- 3. The niche of renal stem cells CD24+CD133+ located in the urinary pole of the Bowman's capsule in humans could contribute to tubular and podocyte regeneration. Stem cells from bone marrow could theoretically differentiate into endothelial and mesangial cells.
- 4. The injury repair process is achieved by slowing the fibrotic response first, and then restoring the tissue architecture of the organ. Administering certain growth factors, at the same time as acting as an antifibrotic, could mobilise stem cells from the kidney and bone marrow. These cells facilitate renal regeneration by fusing with resident renal cells directly, or through paracrine mechanisms.

#### REFERENCES

- Chiurchiu, C, Remuzzi G, Ruggenenti P. Angiotensin-converting enzyme inhibition and renal protection in nondiabetic patients: the data of the meta-analyses. J Am Soc Nephrol 2005;16(Suppl 1):S58-63.[Pubmed]
- Xue JL, et al. Forecast of the number of patients with end-stage renal disease in the United States to the year 2010. J Am Soc Nephrol 2001;12(12):2753-8.[Pubmed]
- Hostetter TH, et al. Hyperfiltration in remnant nephrons: a potentially adverse response to renal ablation. Am J Physiol 1981;241(1):F85-93.[Pubmed]
- Fogo AB. New capillary growth: a contributor to regression of sclerosis? Curr Opin Nephrol Hypertens 2005;14(3):201-3.[Pubmed]
- Feng Z, et al. Glomerular aging in females is a multi-stage reversible process mediated by phenotypic changes in progenitors. Am J Pathol 2005;167(2):355-63.[Pubmed]
- Knoblich JA. Asymmetric cell division during animal development. Nat Rev Mol Cell Biol 2001;2(1):11-20.[Pubmed]
- 7. Hartman HA, Lai HL, Patterson LT. Cessation of renal morphogenesis in mice. Dev Biol 2007;310(2):379-87.[Pubmed]
- Dressler GR. The cellular basis of kidney development. Annu Rev Cell Dev Biol 2006;22:509-29.[Pubmed]
- Ekblom P. Developmentally regulated conversion of mesenchyme to epithelium. FASEB J 1989;3(10):2141-50.[Pubmed]

- Harris RC, Neilson EG. Toward a unified theory of renal progression. Annu Rev Med 2006;57:365-80.[Pubmed]
- 11. Humphreys BD. Slow-cycling cells in renal papilla: stem cells awaken? J Am Soc Nephrol 2009;20(11):2277-9.[Pubmed]
- Maeshima A, Yamashita S, Nojima Y. Identification of renal progenitorlike tubular cells that participate in the regeneration processes of the kidney. J Am Soc Nephrol 2003;14(12):3138-46.[Pubmed]
- Duffield JS, et al. Restoration of tubular epithelial cells during repair of the postischemic kidney occurs independently of bone marrowderived stem cells. J Clin Invest 2005;115(7):1743-55.[Pubmed]
- Lin F, Moran A, Igarashi P. Intrarenal cells, not bone marrowderived cells, are the major source for regeneration in postischemic kidney. J Clin Invest 2005;115(7):1756-64.[Pubmed]
- 15. Oliver JA, et al. The renal papilla is a niche for adult kidney stem cells. J Clin Invest 2004;114(6):795-804.[Pubmed]
- Sagrinati C, et al. Isolation and characterization of multipotent progenitor cells from the Bowman's capsule of adult human kidneys. J Am Soc Nephrol 2006;17(9):2443-56.[Pubmed]
- 17. Humphreys BD, et al. Intrinsic epithelial cells repair the kidney after injury. Cell Stem Cell 2008;2(3):284-91.[Pubmed]
- Romagnani P, Kalluri R. Possible mechanisms of kidney repair. Fibrogenesis Tissue Repair 2009;2(1):3.[Pubmed]
- Ronconi E, et al. Regeneration of glomerular podocytes by human renal progenitors. J Am Soc Nephrol 2009;20(2):322-32.[Pubmed]

- 20. Appel D, et al. Recruitment of podocytes from glomerular parietal epithelial cells. J Am Soc Nephrol 2009;20(2):333-43.[Pubmed]
- Maeshima A. Label-retaining cells in the kidney: origin of regenerating cells after renal ischemia. Clin Exp Nephrol 2007;11(4):269-74.[Pubmed]
- 22. Orlic D, et al. Bone marrow stem cells regenerate infarcted myocardium. Pediatr Transplant 2003;7(Suppl 3):86-8.[Pubmed]
- 23. Ferrari G, et al. Muscle regeneration by bone marrow-derived myogenic progenitors. Science 1998;279(5356):1528-30.[Pubmed]
- 24. Mezey E, et al. Turning blood into brain: cells bearing neuronal antigens generated in vivo from bone marrow. Science 2000;290(5497):1779-82.[Pubmed]
- 25. Lagasse E, et al. Purified hematopoietic stem cells can differentiate into hepatocytes in vivo. Nat Med 2000;6(11):1229-34.[Pubmed]
- 26. Roufosse C, Cook HT. Stem cells and renal regeneration. Nephron Exp Nephrol 2008;109(2):e39-45.[Pubmed]
- Prodromidi EI, et al. Bone marrow-derived cells contribute to podocyte regeneration and amelioration of renal disease in a mouse model of Alport syndrome. Stem Cells 2006;24(11):2448-55.[Pubmed]
- Sugimoto H, et al. Bone-marrow-derived stem cells repair basement membrane collagen defects and reverse genetic kidney disease. Proc Natl Acad Sci USA 2006;103(19):7321-6.
- 29. Zheng F, et al. Development of albuminuria and glomerular lesions in normoglycemic B6 recipients of db/db mice bone marrow: the role of mesangial cell progenitors. Diabetes 2004;53(9):2420-7.[Pubmed]
- 30. Wang X, et al. Cell fusion is the principal source of bone-marrowderived hepatocytes. Nature 2003;422(6934):897-901.[Pubmed]
- Vassilopoulos G, Wang PR, Russell DW. Transplanted bone marrow regenerates liver by cell fusion. Nature 2003;422(6934):901-4.[Pubmed]
- 32. Terada N, et al. Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. Nature 2002;416(6880):542-5.[Pubmed]
- Held PK, et al. In vivo genetic selection of renal proximal tubules. Mol Ther 2006;13(1):49-58.[Pubmed]
- 34. Rafii S, Lyden D. Therapeutic stem and progenitor cell transplantation for organ vascularization and regeneration. Nat Med 2003;9(6):702-12.[Pubmed]
- Anglani F, et al. In search of adult renal stem cells. J Cell Mol Med 2004;8(4):474-87.[Pubmed]
- Morigi M, et al. Mesenchymal stem cells are renotropic, helping to repair the kidney and improve function in acute renal failure. J Am Soc Nephrol 2004;15(7):1794-804.[Pubmed]
- Lange C, et al. Administered mesenchymal stem cells enhance recovery from ischemia/reperfusion-induced acute renal failure in rats. Kidney Int 2005;68(4):1613-7.[Pubmed]
- Duffield JS, Bonventre JV. Kidney tubular epithelium is restored without replacement with bone marrow-derived cells during repair after ischemic injury. Kidney Int 2005;68(5):1956-61.[Pubmed]
- 39. Morigi M, et al. Human bone marrow mesenchymal stem cells accelerate recovery of acute renal injury and prolong survival in mice. Stem Cells 2008;26(8):2075-82.[Pubmed]
- Togel F, et al. Administered mesenchymal stem cells protect against ischemic acute renal failure through differentiation-independent mechanisms. Am J Physiol Renal Physiol 2005;289(1):F31-42.[Pubmed]

- Broekema M, et al. Determinants of tubular bone marrow-derived cell engraftment after renal ischemia/reperfusion in rats. Kidney Int 2005;68(6):2572-81.[Pubmed]
- 42. Bi B, et al. Stromal cells protect against acute tubular injury via an endocrine effect. J Am Soc Nephrol 2007;18(9):2486-96.[Pubmed]
- 43. Karihaloo A, Nickel C, Cantley LG. Signals which build a tubule. Nephron Exp Nephrol 2005;100(1):e40-5.[Pubmed]
- 44. Stuart RO, Nigam SK. Development of the tubular nephron. Semin Nephrol 1995;15(4):315-26.[Pubmed]
- 45. Hammerman MR, Miller SB. Therapeutic use of growth factors in renal failure. J Am Soc Nephrol 1994;5(1):1-11.[Pubmed]
- 46. Nigam S, Lieberthal W. Acute renal failure. III. The role of growth factors in the process of renal regeneration and repair. Am J Physiol Renal Physiol 2000;279(1):F3-F11.[Pubmed]
- 47. Zhang G, et al. A role for fibroblast growth factor type-1 in nephrogenic repair. Autocrine expression in rat kidney proximal tubule epithelial cells in vitro and in the regenerating epithelium following nephrotoxic damage by S-(1,1,2,2-tetrafluoroethyl)-L-cysteine in vivo. J Biol Chem 1993;268(16):11542-7.[Pubmed]
- Shi H, et al. Glial cell line-derived neurotrophic growth factor increases motility and survival of cultured mesenchymal stem cells and ameliorates acute kidney injury. Am J Physiol Renal Physiol 2008;294(1):F229-35.[Pubmed]
- Humes HD, et al. Epidermal growth factor enhances renal tubule cell regeneration and repair and accelerates the recovery of renal function in postischemic acute renal failure. J Clin Invest 1989;84(6):1757-61.[Pubmed]
- 50. Fisher DA, Salido EC, Barajas L. Epidermal growth factor and the kidney. Annu Rev Physiol 1989;51:67-80.[Pubmed]
- Zhuang S, Dang Y, Schnellmann RG. Requirement of the epidermal growth factor receptor in renal epithelial cell proliferation and migration. Am J Physiol Renal Physiol 2004;287(3):F365-72.[Pubmed]
- Baer PC, et al. Expression of a functional epidermal growth factor receptor on human adipose-derived mesenchymal stem cells and its signaling mechanism. Eur J Cell Biol 2009;88(5):273-83.[Pubmed]
- 53. Nakamura T, et al. Molecular cloning and expression of human hepatocyte growth factor. Nature 1989;342(6248):440-3.[Pubmed]
- Miyazawa K, et al. Molecular cloning and sequence analysis of cDNA for human hepatocyte growth factor. Biochem Biophys Res Commun 1989;163(2):967-73.[Pubmed]
- Birchmeier C, Gherardi E. Developmental roles of HGF/SF and its receptor, the c-Met tyrosine kinase. Trends Cell Biol 1998;8(10):404-10.[Pubmed]
- 56. Xiao GH, et al. Anti-apoptotic signaling by hepatocyte growth factor/Met via the phosphatidylinositol 3-kinase/Akt and mitogenactivated protein kinase pathways. Proc Natl Acad Sci USA 2001;98(1):247-52.
- 57. Mizuno S, et al. Reciprocal balance of hepatocyte growth factor and transforming growth factor-beta 1 in renal fibrosis in mice. Kidney Int 2000;57(3):937-48.[Pubmed]
- Dai C, Liu Y. Hepatocyte growth factor antagonizes the profibrotic action of TGF-beta1 in mesangial cells by stabilizing Smad transcriptional corepressor TGIF. J Am Soc Nephrol 2004;15(6):1402-12.[Pubmed]
- 59. Inoue T, et al. TGF-beta1 and HGF coordinately facilitate collagen turnover in subepithelial mesenchyme. Biochem Biophys Res Commun 2002;297(2):255-60.[Pubmed]

- Mizuno S, et al. Hepatocyte growth factor prevents renal fibrosis and dysfunction in a mouse model of chronic renal disease. J Clin Invest 1998;101(9):1827-34.[Pubmed]
- 61. Ueki T, et al. Hepatocyte growth factor gene therapy of liver cirrhosis in rats. Nat Med 1999;5(2):226-30.[Pubmed]
- 62. Gao X, et al. Hepatocyte growth factor gene therapy retards the progression of chronic obstructive nephropathy. Kidney Int 2002;62(4):1238-48.[Pubmed]
- 63. Yang R, et al. Hemodynamic effects of scatter factor in conscious rats. J Cardiovasc Pharmacol 1997;30(3):294-301.[Pubmed]
- 64. Kollet O, et al. HGF, SDF-1, and MMP-9 are involved in stressinduced human CD34 stem cell recruitment to the liver. J Clin Invest 2003;112(2):160-9.[Pubmed]
- Higashiyama R, et al. Bone marrow-derived cells express matrix metalloproteinases and contribute to regression of liver fibrosis in mice. Hepatology 2007;45(1):213-22.[Pubmed]
- 66. Huh CG, et al. Hepatocyte growth factor/c-met signaling pathway is required for efficient liver regeneration and repair. Proc Natl Acad Sci USA 2004;101(13):4477-82.
- 67. Mayer H, et al. Vascular endothelial growth factor (VEGF-A) expression in human mesenchymal stem cells: autocrine and

paracrine role on osteoblastic and endothelial differentiation. J Cell Biochem 2005;95(4):827-39.[Pubmed]

- 68. Geiger F, et al. VEGF producing bone marrow stromal cells (BMSC) enhance vascularization and resorption of a natural coral bone substitute. Bone 2007 41(4):516-22.
- 69. Hung SC, et al. Angiogenic effects of human multipotent stromal cell conditioned medium activate the PI3K-Akt pathway in hypoxic endothelial cells to inhibit apoptosis, increase survival, and stimulate angiogenesis. Stem Cells 2007;25(9):2363-70. [Pubmed]
- Hohenstein B, et al. Enhanced progenitor cell recruitment and endothelial repair after selective endothelial injury of the mouse kidney. Am J Physiol Renal Physiol 2010;298(6):F1504-14.[Pubmed]
- 71. Imasawa T, et al. The potential of bone marrow-derived cells to differentiate to glomerular mesangial cells. J Am Soc Nephrol 2001;12(7):1401-9.[Pubmed]
- 72. Masuya M, et al. Hematopoietic origin of glomerular mesangial cells. Blood 2003;101(6):2215-8.[Pubmed]
- Rookmaaker MB, et al. Bone-marrow-derived cells contribute to glomerular endothelial repair in experimental glomerulonephritis. Am J Pathol 2003;163(2):553-62.[Pubmed]