

Case report

Autosomal dominant distal renal tubular acidosis in two pediatric patients with mutations in the SLC4A1 gene. Can the maximum urinary pCO₂ test be normal?

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ABSTRACT

Primary distal renal tubular acidosis (dRTA) is a rare tubulopathy characterised by the presence of hyperchloremic metabolic acidosis. It is caused by the existence of a defect in the function of the H⁺-ATPase located on the luminal side of the α-intercalated cells or the Cl⁻-HCO₃⁻ (AE1) anion exchanger located on the basolateral side. Patients do not acidify the urine after acid overload (NH₄Cl) or after stimulating H⁺ secretion by obtaining a high intratubular concentration of an anion such as chlorine (pH is measured) or HCO₃⁻ (urinary pCO₂ is measured). We present a family with autosomal dominant dRTA produced by a heterozygous mutation in the SLC4A1 gene in which the two paediatric members showed a test of normal maximum urinary pCO₂. Our hypothesis is that since the H⁺-ATPase is intact, at least initially, the stimulation induced by intratubular electronegativity to secrete H⁺ could be effective, which would allow the maximum urinary pCO₂ to be paradoxically normal, which could explain the onset, moderate presentation of symptoms and late diagnosis in patients with this mutation. This is the first documented case of a dominant dRTA in Mexico.

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Acidosis tubular renal distal autosómica dominante en dos pacientes pediátricos con mutaciones en el gen SLC4A1. ¿La prueba de la pCO₂ urinaria máxima puede ser normal?

RESUMEN

Palabras clave:

Acidosis tubular renal
Nefrocalcinosis
Litiasis
pCO₂ urinaria máxima

La acidosis tubular renal distal (ATRd) primaria es una tubulopatía poco frecuente caracterizada por la presencia de acidosis metabólica hiperclorémica. Está generada por la existencia de un defecto en la función de la H⁺-ATPasa situada en el lado luminal de las células α -intercaladas o del intercambiador de aniones Cl⁻-HCO₃⁻ (AE1) ubicado en el lado basolateral. Los pacientes no acidifican la orina tras una sobrecarga ácida (NH₄Cl) o tras estimular la secreción de H⁺ mediante la obtención de una elevada concentración intratubular de un anión como cloro (se mide el pH) o HCO₃⁻ (se mide la pCO₂ urinaria). Se presenta una familia con ATRd autosómica dominante producida por una mutación heterocigota en el gen SLC4A1 en la que los dos miembros en edad pediátrica mostraron una prueba de la pCO₂ urinaria máxima normal. Nuestra hipótesis es que al estar intacta, al menos inicialmente, la H⁺-ATPasa, podría ser efectivo el estímulo inducido por la electronegatividad intratubular para secretar H⁺ lo que permitiría que la pCO₂ urinaria máxima fuera paradójicamente normal, lo que pudiera explicar el inicio tardío, la presentación moderada de los síntomas y el diagnóstico en edades más avanzadas, en los pacientes con dicha mutación. Este es el primer caso documentado de una ATRd dominante en México.

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Introduction

Distal renal tubular acidosis (dRTA) (*Butler-Albright disease, nephrocalcinosis infantum*) is a clinical syndrome characterised by hyperchloraemic metabolic acidosis. It is caused by a defect in the urinary excretion of hydrogen ion (H⁺) in the distal portions of the nephron, which makes them unable to lower the urine pH below 6.^{1,2} It is a rare disease, which causes growth retardation, polyuria, hypercalciuria, hypocitraturia, nephrocalcinosis, potassium depletion and characteristic extrarenal manifestations in some of the types.^{2,3}

Distal tubular acidification of urine under normal conditions is summarised in Fig. 1. Vacuolar H⁺-ATPases (V-ATPase) are highly conserved proton pumps expressed in α -intercalated cells and are made up of two domains, V1 and V0. Defects in the activity of the H⁺-ATPase pump cause the majority of primary cases of autosomal recessive hereditary dRTA. They are due to mutations in the ATP6V1B1 and ATP6V0A4 genes (Table 1) that encode the B1 and A4 subunits of the H⁺-ATPase pump, respectively.^{4–6} Autosomal recessive dRTA associated with deafness can be caused by mutations in the FOXI1 gene that encodes a transcription factor necessary for the expression of at least four subunits of H⁺-ATPase (A1, B1, E2 and a4).⁷ A fourth gene, WDR72 (OMIM 613211), when mutated produces another variety of autosomal recessive dRTA. This gene seems to be involved in the intracellular trafficking of proteins that regulate the acid-base balance, causing their intracellular retention or misdirection.^{8,9} Extrarenal manifestations include: 1) patients with recessive dRTA caused by mutations in the ATP6V1B1 or ATP6V0A4 genes may frequently and constantly have sensorineural deafness in FOXI1 gene mutations, with early onset and associated with Pendred

syndrome (ORPHA:705);⁸ and 2) patients with recessive dRTA caused by WDR72 gene mutations may develop amelogenesis imperfecta.

Basolaterally, only one type of autosomal dominant inheritance of dRTA has been described (Table 1) in most cases. It is caused by mutations in the SLC4A1 (OMIM 109270) gene, which encodes the two isoforms of the Cl⁻-HCO₃⁻ anion exchanger, the renal isoform also known as kAE1 (kidney anion exchanger 1) and the red blood cell isoform also known as band 3 protein (erythroid isoform or eAE1); kAE1 is responsible for HCO₃⁻ reabsorption along with Cl⁻ excretion in α -intercalated cells (Fig. 1).¹⁰ This type of dRTA has been associated with less severe forms of clinical presentation, with late onset in childhood, adolescence and in adult patients, with less impact on growth compared to autosomal recessive inheritance.⁸

It has been reported that patients with dRTA do not acidify their urine after an acid overload (NH₄Cl) or after H⁺ secretion being stimulated by a high intratubular concentration due to an anion such as chlorine (furosemide, Cl₂Ca). Neither does their urinary pCO₂ increased above 70 mmHg despite a high intratubular concentration of HCO₃⁻.¹¹ We present a family with autosomal dominant dRTA caused by a heterozygous mutation in the SLC4A1 gene, in which two of its paediatric members had a normal maximum urinary pCO₂ test.

Case reports

Case 1

This was a female patient aged four years three months. Birth weight 2800 g, length 48 cm and Apgar 8/9. History of craving for water and salt. Polyuria and nocturia as an infant. She

Table 1 – OMIM hereditary distal renal tubular acidosis.

Gene	Protein	Disease OMIM	Gene OMIM	Transmission
SLC4A1	Exchanger (AE1)	#179800	*109270	AD, AR
ATP6V1B1	Subunit B1 of the H ⁺ -ATPase pump (V-ATPase)	#267300	*192132	AR
ATP6VOA4	Subunit a4 of the H ⁺ -ATPase pump (V-ATPase)	#602722	*605239	AR

Table 2 – Blood and urine clinical chemistry tests.

	Case 1	Case 2	Case 3
Cystatin C (mg/l)	0.91	0.99	
eGFR (ml/min/1.73 m ²) ^a	102.03	92.12	122
Venous blood gases			
pH	7.33	7.04	7.17
HCO ₃ ⁻ (mEq/l)	14.8	16.7	14.1
Blood chloride (mmol/l)	112	114	113
Blood potassium (mmol/l)	2.5	3.5	3.3
Blood anion gap	11.2	8.3	10.9
Blood uric acid (mg/dl)	1.3	1.5	3.3
Urinary anion gap	25.8	55.4	36
Urinary calcium (mg/kg/day)	10.5	8.9	
Urinary calcium/creatinine ratio (mg/mg)	0.95	0.84	
Urinary citrate (mg/kg/day)	0.5	0.42	0.24
Urinary citrate/creatinine ratio (mg/g)	0.413	—	
Urinary calcium/citrate ratio (mg/mg)	21	21.19	
FE HCO ₃ ⁻ (ml/100 ml GFR)	7	5.2	
FEK (ml/100 ml GFR)	29.5	17	
FE uric acid (ml/100 ml GFR)	16	13.9	
Urinary protein (mg/m ² /h)	38	62	

eGFR: estimated renal glomerular filtration rate; FE: fractional excretion; FEK: fractional excretion of potassium; GFR: renal glomerular filtration rate; HCO₃⁻: bicarbonate anion.

^a Berg's formula (2015).

was hospitalised 11 times for fever with no obvious focus. At the age of 20 months, she was diagnosed with herpes zoster and treated with aciclovir for seven days; subsequently, she developed diarrhoea, moderate dehydration and general flaccidity with the inability to walk. She was admitted to the Hospital General de Zona with suspected Guillain-Barré syndrome; weight 9.5 kg and height 82 cm (both in the 10th percentile). She was found to have hypokalaemia (2.5 mEq/l) and metabolic acidosis. After being started on intravenous treatment and the correction of her fluid/electrolyte imbalance, her strength improved and she was able to walk. However, the hyperchloraemic metabolic acidosis persisted. The clinical chemistry data are shown in **Table 2** and the ultrasound findings in **Fig. 2**. A hearing test showed no evidence of sensorineural hearing loss. Her ophthalmological examination was normal. A urinary acidification test with sodium bicarbonate and acetazolamide was performed according to a previously described protocol (maximum urinary pCO₂ test).^{11–13} The test was without complications, well tolerated and considered valid when bicarbonaturia greater than 80 mEq/l was found. A maximum urinary pCO₂ of 80 mmHg was measured at 60 min (abnormal <70 mmHg) (**Table 3**). The patient was found to be a carrier of the same mutation as her mother. After three years of treatment with alkalis, her weight and height have improved (16.5 kg [47th percentile] and 103 cm [34th percentile], respectively). She is currently on treatment with potassium citrate (4.5 mEq/kg/day) and her hypercalciuria has remitted. Her blood pressure (BP) is normal (89/59 mmHg).

Table 3 – Maximum urinary pCO₂ test performed after stimulation with sodium bicarbonate and acetazolamide.

	Case 1	Case 2
Urinary pH	7.9	7.9
Maximum urinary pCO ₂ (mmHg)	80	70
Urinary HCO ₃ ⁻ (mEq/l)	153.37	126.18

HCO₃⁻: bicarbonate anion; pCO₂: partial pressure of carbon dioxide.

Case 2

This was a male patient aged eight years 11 months with no relevant perinatal history (weight 2.9 kg, length 50 cm and Apgar 9/10). Cravings for water and salt from about two years of age. He had to be admitted to hospital three times before the age of 12 months with episodes of moderate dehydration. He was taken to the local Accident and Emergency department 24 times with fever. On some occasions it was classified as a respiratory tract infection, but in most cases no apparent cause was identified. He was investigated after his sister was diagnosed with dRTA. He weighed 18 kg (5th percentile) and his height was 111 cm (3rd percentile). He was found to have hyperchloraemic metabolic acidosis with normal anion gap, hypercalciuria, severe hypocitraturia (**Table 2**) and grade III nephrocalcinosis. His hearing test was normal. Maximum urinary pCO₂ was also normal (**Table 3**). Both his weight (23 kg, 8th percentile) and his height (122 cm, 17th percentile) have also improved after three years of treatment with alkalis. He is on

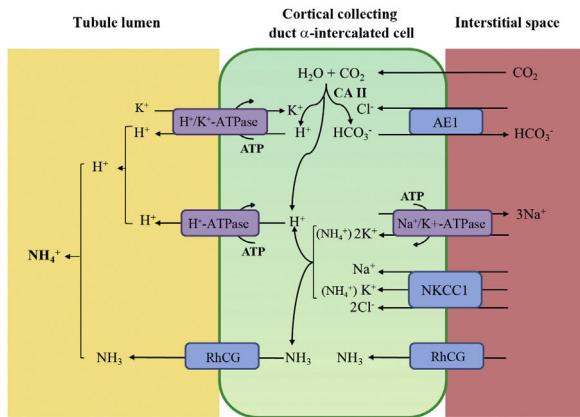


Fig. 1 – Distal urine acidification under normal conditions. Urine acidification occurs in the distal and collecting tubules through three related processes: a) reabsorption of the residual amount of bicarbonate (10%), which has not been recovered in more proximal areas of the nephron; b) titration of the divalent phosphate anion (HPO_4^{2-}) with H^+ , which is transformed into monovalent phosphate anion (H_2PO_4^-) or titratable acidity; c) accumulation of intraluminal ammonia (NH_3), which take up H^+ and forms ammonium (NH_4^+). The secretion of H^+ and the titration of urinary buffers lead to acidification of the urine, with which pH values close to 4.5 can be reached under conditions of maximum stimulation of the process. This takes place in the α -intercalated cells in the late distal convoluted tubule, the connecting tubule and in the cortical and medullary collecting ducts.

Distal secretion of H^+ generates an equimolar amount of bicarbonate. For each hydrogen molecule excreted into the tubular lumen, a new bicarbonate molecule is generated intracellularly thanks to the action of intracytoplasmic carbonic anhydrase (CA type II), which is transferred to the blood through the Cl^- - HCO_3^- anion exchanger (AE1). The α -intercalated cells secrete H^+ via vacuolar ATPase (H^+ -ATPase), which actively transfers H^+ across the luminal membrane, and H^+ - K^+ -ATPase, which exchanges H^+ for potassium. The function of H^+ -ATPase is markedly influenced by the electronegativity generated in the tubular lumen by the simultaneous transport of Na^+ in the principal cells of the collecting duct. The accumulation of $\text{NH}_3/\text{NH}_4^+$ in the medulla generates a concentration gradient that helps it to cross through the basolateral membrane of the α -intercalated cells. $\text{NH}_3/\text{NH}_4^+$ excretion requires at least two steps: basolateral entry and luminal excretion. Uptake from the $\text{NH}_3/\text{NH}_4^+$ interstitium is carried out by several pathways including the $\text{Na}^+/\text{K}^+/2\text{Cl}^-$ transporter (NKCC1), Na^+/K^+ -ATPase (in the case of these two transporters, NH_4^+ can be transported instead of potassium), RhCG (human Rhesus C glycoprotein) gas channels and hyperpolarisation-activated cyclic nucleotide-gated (HCN) cationic non-selective NH_4^+ channels. The luminal membrane has a high permeability for NH_3 . RhCG channels are present in the luminal membrane and in the basolateral membrane.



Fig. 2 – Renal ultrasound of case 1, with pyramid-shaped hyperechogenic images diffusely distributed in the medullary region without projecting a posterior acoustic shadow. Consistent with severe bilateral nephrocalcinosis.

treatment with potassium citrate (5 mEq/kg/day). The genetic study showed the same mutation as his mother and sister.

In both children the morphology of their red blood cells was normal.

Case 3

This was the 29-year-old mother of the patients, with a history of mother with diabetes and father with hypertension on treatment. She had an older brother with kidney stones and multiple episodes of flaccid paralysis. She was diagnosed after the investigations on her children. She had had a craving for water since childhood. She weighed 58 kg, was 157 cm tall and her blood pressure was 100/60 mmHg. Creatinine levels were 0.78 mg/dl (CrCl 122 ml/min), sodium 139 mEq/l, potassium 3.3 mEq/l and blood chloride 113 mEq/l. Venous blood gas analysis revealed metabolic acidosis (pH 7.17, HCO_3^- 14.1 mEq/l) with hypocitraturia (14 mg/24 h) and proteinuria 382 mg/24 h (Table 2). Her urinary pH was 7 and she had a positive urine anion gap (+36). Renal ultrasound revealed grade II-III medullary nephrocalcinosis (Fig. 3). A calculus was found in the lower segment of her right ureter and another 1.5 cm in size in her bladder. A genetic study was performed, analysing the genes ATP6VOA4, ATP6V1B1 and SLC4A1. A heterozygous pathogenic variant was found in exon 20 of the gene SLC4A1, c.2710_*12, p. (Tyr904_Val911delins68), resulting in a deletion of 39 bases (27 from the last exon and 12 from the 3' UTR region), modifying the carboxy terminus of the protein (deletion of the last 8 amino acids and insertion of a new sequence of 68 amino acids). This variation, which is not described in the literature and not included in the database, is considered pathogenic, class 5 according to the American College of Medical Genetics and Genomics (ACMG) 2015 guidelines; the criteria used to classify it as class 5 are: PVS1, PM2, PM1 and PM4. This variation is not in the general population Genome Aggregation Database (gnomAD) (criterion PM2).

The peripheral blood samples were collected in EDTA tubes. Pure DNA was obtained using the QIAamp® DNA Blood MIDI Kit (Qiagen) in accordance with the manufacturer's instructions. The index case of this family was analysed with the gene panel using next-generation sequencing (previously

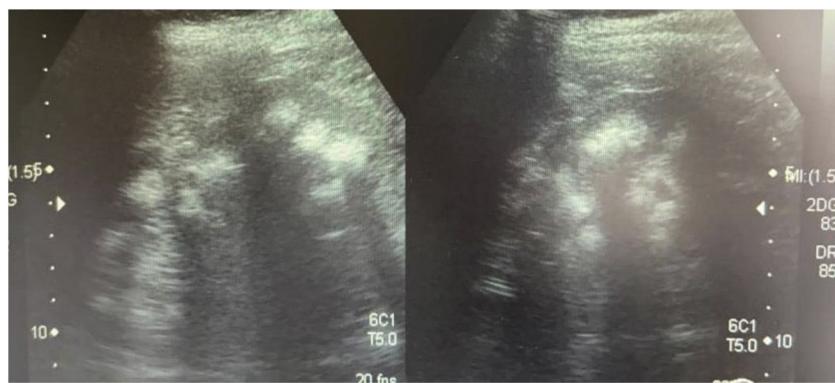


Fig. 3 – Ultrasound performed on the mother of the patients. Kidneys with heterogeneous echogenicity due to increased echogenicity of the renal pyramids consistent with grade II-III bilateral medullary nephrocalcinosis. Corticomedullary differentiation is poor, with signs of diffuse chronic nephropathy.

described in Ashton E et al., *Kidney Int*, 2018 PMID: 29398133 and Hureaux M, *Kidney Int*, 2019 PMID: 31672324), which enabled analysis of the three major genes; FOXI1 was not analysed as it was not included in the panel at the time the study was conducted. Once the variation was identified, it was confirmed in the index case using the Sanger method and the other members of the family were also analysed with Sanger. The transcripts used to describe the variations were: ATP6V0A4: NM_130841.2; ATP6V1B1: NM_001692.3; and SLC4A1: NM_000342.3.

Discussion

When there is a genetic defect in the activity of the H^+ -ATPase pump, the metabolic acidosis caused by the defect in tubular secretion of H^+ causes growth retardation, mobilisation of bone calcium¹⁴ and inhibition of tubular reabsorption of $NaCl$ and calcium.¹⁵ Citrate is a very sensitive marker of metabolic acidosis.¹⁶ Reduction in the pH of the proximal tubular cell stimulates the activity of the dicarboxylate cotransporter, sodium-dependent dicarboxylate transporter 1 (NADC1), located in the apical membrane of the proximal tubule. NADC1 is a Na^+ -coupled electrogenic transporter mechanism for reabsorption (3 Na^+ cations for every Cit^{2-} anion). The consequence of increased tubular reabsorption is hypocitraturia. Once the citrate is reabsorbed, it is transported to the mitochondria where mitochondrial aconitase transforms it into isocitrate to enter the Krebs cycle¹⁷ in which citrate is converted to CO_2 and H_2O and three H^+ are consumed, a reaction equivalent to generating bicarbonate. Hypercalciuria and hypocitraturia can increase the risk of nephrocalcinosis. As Na^+ cannot be exchanged for H^+ in the distal portions of the nephron, it needs to be exchanged with K^+ . Hypokalaemia can be a consequence of secondary hyperaldosteronism caused by volume contraction, which, in turn, is caused by salt loss and polyuria. Polyuria is the result of nephrocalcinosis, the aforementioned salt loss, body potassium depletion (hypokalaemic nephropathy)^{18,19} and, certainly, hypercalciuria itself.

What happens when the genetic defect affects the activity of the Cl^- - HCO_3^- anion exchanger, as in our patients? Bicarbonate, formed intracellularly by the action of carbonic anhydrase, cannot leave the cell through the basolateral membrane, so the intracellular pH rises, which inhibits the activity of H^+ -ATPase. Patients with autosomal dominant dRTA therefore have urine with a pH above 6 in a situation of spontaneous acidosis²⁰ or in acidification tests carried out with H^+ stimulation, as in those performed with NH_4Cl .^{20,21} The lack of exchange with Cl^- explains the hyperchloraemia typical of dRTA (Fig. 1).

However, as the H^+ -ATPase is intact, at least initially, the stimulus induced by intratubular electronegativity may be effective. Since the 1980s, it has been recognised that distal acidification capacity is dependent on anions²² and is stimulated by aldosterone.²³ That explains at least two important points. Firstly, that clinical manifestation is later in patients with the autosomal dominant form, especially in childhood and adolescence, and is commonly a causal finding in adults; with a milder phenotype in adolescence or adulthood, with slightly low serum bicarbonate levels and with less involvement and compromise of weight and height, which tend to remain within appropriate percentiles for age.^{3,24} Some patients with heterozygous mutations in the SLC4A1 gene even have an incomplete dRTA phenotype, that is, without metabolic acidosis^{20,25,26} (in some patients with incomplete dRTA, the pCO_2 test is normal²⁵). In the course of evolution, it seems that the transcription of the different subunits that make up H^+ -ATPase must be progressively altered, causing the symptoms to become more evident. Secondly, the electronegativity caused by the high intratubular concentration of bicarbonate would stimulate H^+ -ATPase to secrete H^+ , which would result in normal maximum urinary pCO_2 test results, as we found in our two patients.

Another important finding in the three cases we report here is hypouricaemia associated with hyperuricosuria, which has been described particularly in autosomal recessive forms in some cases of dRTA, as a consequence of endosomal dysfunction of proximal tubule cells induced by chronic intracellular

acidosis or associated with nephropathy secondary to chronic hypokalaemia, which causes vacuolisation of proximal tubule cells.^{27,28}

The very significant nephrocalcinosis patients develop is particularly striking. This has been reported in 94% of dRTA cases with mutations in the SLC4A1 gene, and is slightly more common, at 98%, in ATP6V0A4 mutations,²⁴ most likely due to the severe hypocitraturia in our three cases, as well as the elevated hypercalciuria they had at diagnosis, which returned to normal once treatment potassium citrate was started.

The inheritance of this type of dRTA with mutations in the SLC4A1 gene is complex with, as mentioned above, autosomal dominant transmission, especially in Caucasians.²¹ However, a recessive form has also been described, which is more common in some tropical countries of Southeast Asia, particularly in Thailand, Malaysia, the Philippines and Papua New Guinea; in these countries, ovalocytosis due to heterozygous mutations of SLC4A1 (Southeast Asian ovalocytosis [SAO]) is common.^{29,30} Other mutations in this gene can also cause spherocytosis. Mutations that cause ovalocytosis or spherocytosis are responsible for morphological changes in red blood cells and haemolytic anaemia, and affect amino acids different from those affected in dRTA. Walsh et al.³¹ and other groups³² have tried to identify why dRTA is so common in Southeast Asia. Their published results suggest the hypothesis that the changes in red blood cells caused by these mutations could protect against malaria. In 1981, oval red blood cells from Melanesians living in Papua New Guinea were shown to be resistant to infection by malaria parasites (*Plasmodium falciparum*).³³ Hereditary spherocytosis is a phenotypic and genetically heterogeneous condition. Some 20%–35% of cases are caused by mutations in the SLC4A1 gene.³⁴ There have been reports of patients with biallelic mutations in this gene (for example, an SAO mutation and a distal acidosis mutation) who may suffer from haemolytic anaemia associated with dRTA.^{35–37}

In 1999, Kaitwatcharachai et al. published the data of an adult patient with SAO in whom the urinary pCO₂ test was normal.³⁸ Years later, one of the co-authors of that paper reflected on the finding and suggested that some of the mutated kAE1 proteins would be expressed in the luminal membrane, which would produce “a net secretion of bicarbonate and an increase in pCO₂ in the distal nephron”.³⁹ To our knowledge, this hypothesis would not explain the increase in H⁺ secretion necessary to increase urinary pCO₂.

In any event, the heterozygous mutation in exon 20 of the SLC4A1 gene found in our patients is the first time it has been described. Therefore, the return of urinary pCO₂ to normal levels seen in our patients should not be specific to patients carrying the SAO mutation.

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Conflicts of interest

The authors declare that they have no conflicts of interest.

REFERENCES

1. Butler AM, Wilson JL, Farber S. Dehydration and acidosis with calcification at renal tubules. *J Pediatr*. 1936;8:489–99.
2. Rodríguez Soriano J. Renal tubular acidosis: the clinical entity. *J Am Soc Nephrol*. 2002;13:2160–70, <http://dx.doi.org/10.1097/01.asn.0000023430.92674.e5>.
3. Gil-Peña H, Mejía N, Santos F. Renal tubular acidosis. *J Pediatr*. 2014;164:691–8, <http://dx.doi.org/10.1016/j.jpeds.2013.10.085>.
4. Karet FE, Finberg KE, Nelson RD, Nayir A, Mocan H, Sanjad SA, et al. Mutations in the gene encoding B1 subunit of H⁺-ATPase cause renal tubular acidosis with sensorineural deafness. *Nat Genet*. 1999;21:84–90, <http://dx.doi.org/10.1038/5022>.
5. Smith AN, Skaug J, Choate KA, Nayir A, Bakkaloglu A, Ozen S, et al. Mutations in ATP6N1B, encoding a new kidney vacuolar proton pump 116-kD subunit, cause recessive distal renal tubular acidosis with preserved hearing. *Nat Genet*. 2000;26:71–5, <http://dx.doi.org/10.1038/79208>.
6. Jobst-Schwan T, Klämbt V, Tarsio M, Heneghan JF, Majmundar AJ, Shril S, et al. Whole exome sequencing identified ATP6V1C2 as a novel candidate gene for recessive distal renal tubular acidosis. *Kidney Int*. 2020;97:567–79, <http://dx.doi.org/10.1016/j.kint.2019.09.026>.
7. Enerbäck S, Nilsson D, Edwards N, Heglind M, Alkanderi S, Ashton E, et al. Acidosis and deafness in patients with recessive mutations in FOXI1. *J Am Soc Nephrol*. 2018;29:1041–8, <http://dx.doi.org/10.1681/ASN.2017080840>.
8. Alexander RT, Law L, Gil-Peña A, Greenbaum LA, Santos F, Adamet MP, et al. Hereditary distal renal tubular acidosis. In: Adam MP, Arlinger HH, Pagon RA, Wallace SE, Bean LJH, Mirzaa G, Amemiya A, editors. *GeneReviews®*. Seattle (WA): University of Washington, Seattle; 2019. p. 1993–2021.
9. Rungraj N, Nettuwakul C, Sawasdee N, Sangnual S, Deejai N, Misgar RA, et al. Distal renal tubular acidosis caused by tryptophan-aspartate repeat domain 72 (WDR72) mutations. *Clin Genet*. 2018;94:409–18, <http://dx.doi.org/10.1111/cge.13418>.
10. Wagner S, Vogel R, Lietzke R, Koob R, Drenckhahn D. Immunochemical characterization of a band 3-like anion exchanger in collecting duct of human kidney. *Am J Physiol*. 1987;253:F213–21, <http://dx.doi.org/10.1152/ajprenal.1987.253.2.F213>.
11. Halperin ML, Goldstein MB, Haig A, Johnson MD, Stinebaugh BJ. Studies on the pathogenesis of type I (distal) renal tubular acidosis as revealed by the urinary pCO₂ tensions. *J Clin Invest*. 1974;53:669–77, <http://dx.doi.org/10.1172/JCI107604>.
12. García Nieto V, Hernández-González MJ, Hernández-Hernández L, Monge M, Molini N. A new combined test to study the maximum urinary pCO₂ in the pediatric age. *Pediatr Nephrol*. 2002;17: C8 (Abstract).
13. Guerra-Hernandez NE, Ordaz-Lopez KV, Escobar-Perez L, Gomez-Tenorio C, Garcia-Nieto VM. Distal renal tubular acidosis screening by urinary acidification testing in Mexican children. *Rev Invest Clin*. 2015;67:191–8.
14. Lemann JJ Jr, Litzow JR, Lennon EJ. The effects of chronic acid loads in normal man: further evidence for the participation of bone mineral in the defense against chronic metabolic acidosis. *J Clin Invest*. 1966;45:1608–14, <http://dx.doi.org/10.1172/JCI105467>.
15. Mahnensmith R, Thier SO, Cooke CR, Broadus A, DeFronzo RA. Effect of acute metabolic acidemia on renal electrolyte

- transport in man. *Metabolism.* 1979;28:831–42, [http://dx.doi.org/10.1016/0026-0495\(79\)90210-5](http://dx.doi.org/10.1016/0026-0495(79)90210-5).
16. Brennan S, Hering-Smith K, Hamm LL. Effect of pH on citrate reabsorption in the proximal convolute tubule. *Am J Physiol.* 1988;255:F301–6, <http://dx.doi.org/10.1152/ajprenal.1988.255.2.F301>.
 17. Tosukhowong P, Tungsanga K, Phongudom S, Sriboonlue P. Effects of potassium-magnesium citrate supplementation on cytosolic ATP citrate lyase and mitochondrial aconitase activity in leukocytes: a window on renal citrate metabolism. *Int J Urol.* 2005;12:140–4, <http://dx.doi.org/10.1111/j.1442-2042.2005.01001.x>.
 18. Manitius A, Levitin H, Beck D, Epstein FH. On the mechanism of impairment of renal concentrating ability in potassium deficiency. *J Clin Invest.* 1960;39:684–92, <http://dx.doi.org/10.1172/JCI104084>.
 19. Emery G, Young RM, Morgan DB, Hay AW, Tete-Donker D, Rubython J. Tubular damage in patients with hypokalaemia. *Clin Chim Acta.* 1984;140:231–8, [http://dx.doi.org/10.1016/0009-8981\(84\)90204-3](http://dx.doi.org/10.1016/0009-8981(84)90204-3).
 20. Bruce LJ, Cope DL, Jones GK, Schofield AE, Burley M, Povey S, Unwin RJ, Wrong O, Tanner MJ. Familial distal renal tubular acidosis is associated with mutations in the red cell anion exchanger (Band 3, AE1) gene. *J Clin Invest.* 1997;100:1693–707, <http://dx.doi.org/10.1172/JCI119694>.
 21. Karet FE, Gainza FJ, Györy AZ, Unwin RJ, Wrong O, Tanner MJ, et al. Mutations in the chloride-bicarbonate exchanger gene AE1 cause autosomal dominant but not autosomal recessive distal renal tubular acidosis. *Proc Natl Acad Sci U S A.* 1998;95:6337–42, <http://dx.doi.org/10.1073/pnas.95.11.6337>.
 22. Stone DK, Seldin DW, Kokko JP, Jacobson HR. Anion dependence of rabbit medullary collecting duct acidification. *J Clin Invest.* 1983;71:1505–8, <http://dx.doi.org/10.1172/jci110905>.
 23. Stone DK, Seldin DW, Kokko JP, Jacobson HR. Mineralocorticoid modulation of rabbit medullary collecting duct acidification. A sodium-independent effect. *J Clin Invest.* 1983;72:77–83, <http://dx.doi.org/10.1172/jci110986>.
 24. López-García SC, Emma F, Walsh SB, Fila M, Hooman N, Zaniew M, et al. Treatment and long-term outcome in primary distal renal tubular acidosis. *Nephrol Dial Transplant.* 2019;34:981–91, <http://dx.doi.org/10.1093/ndt/gfy409>.
 25. Rysavá R, Tesar V, Jirsa M Jr, Brabec V, Jarolím P. Incomplete distal renal tubular acidosis coinherited with a mutation in the band 3 (AE1) gene. *Nephrol Dial Transplant.* 1997;12:1869–73, <http://dx.doi.org/10.1093/ndt/12.9.1869>.
 26. Takeuchi T, Hattori-Kato M, Okuno Y, Kanatani A, Zaitsu M, Mikami K. A single nucleotide polymorphism in kidney anion exchanger 1 gene is associated with incomplete type 1 renal tubular acidosis. *Sci Rep.* 2016;6:35841, <http://dx.doi.org/10.1038/srep35841>.
 27. Watanabe T. Proximal renal tubular dysfunction in primary distal renal tubular acidosis. *Pediatr Nephrol.* 2005;20:86–8, <http://dx.doi.org/10.1007/s00467-004-1693-8>.
 28. Tassic V. Atypical presentation of distal renal tubular acidosis in two siblings. *Pediatr Nephrol.* 2008;23:1177–81, <http://dx.doi.org/10.1007/s00467-008-0796-z>.
 29. Nilwarangkur S, Nimmannit S, Chaovakul V, Susaengrat W, Ong-aj-Yooth S, Vasuvattakul S, et al. Endemic primary distal renal tubular acidosis in Thailand. *Q J Med.* 1990;74:289–301.
 30. Yenchitsomanus PT, Vasuvattakul S, Kirdpon S, Wasanawatana S, Susaengrat W, Srerethiphayawan S, et al. Autosomal recessive distal renal tubular acidosis caused by G701D mutation of anion exchanger 1 gene. *Am J Kidney Dis.* 2002;40:21–9, <http://dx.doi.org/10.1053/ajkd.2002.33909>.
 31. Khositseth S, Bruce LJ, Walsh SB, Bawazir WM, Ogle GD, Unwin RJ, et al. Tropical distal renal tubular acidosis: clinical and epidemiological studies in 78 patients. *Q J Med.* 2012;105:861–77, <http://dx.doi.org/10.1093/qjmed/hcs139>.
 32. Walsh S, Borgese F, Gabillat N, Guizouarn H. Southeast Asian AE1 associated renal tubular acidosis: cation leak is a class effect. *Biochem Biophys Res Commun.* 2009;382:668–72, <http://dx.doi.org/10.1016/j.bbrc.2009.03.062>.
 33. Kidson C, Lamont G, Saul A, Nurse GT. Ovalocytic erythrocytes from Melanesians are resistant to invasion by malaria parasites in culture. *Proc Natl Acad Sci U S A.* 1981;78:5829–32, <http://dx.doi.org/10.1073/pnas.78.9.5829>.
 34. van Vuren A, van der Zwaag B, Huisjes R, Lak N, Bierings M, Gerritsen E, et al. The complexity of genotype-phenotype correlations in hereditary spherocytosis: a cohort of 95 patients: genotype-phenotype correlation in hereditary spherocytosis. *Hemasphere.* 2019;3:e276, <http://dx.doi.org/10.1097/HS9.0000000000000276>.
 35. Ribeiro ML, Alloisio N, Almeida H, Gomes C, Texier P, Lemos C, et al. Severe hereditary spherocytosis and distal renal tubular acidosis associated with the total absence of band 3. *Blood.* 2000;96:1602–4.
 36. Chang YH, Shaw CF, Jian SH, Hsieh KH, Chiou YH, Lu PJ. Compound mutations in human anion exchanger 1 are associated with complete distal renal tubular acidosis and hereditary spherocytosis. *Kidney Int.* 2009;76:774–83, <http://dx.doi.org/10.1038/ki.2009.258>.
 37. Chu C, Woods N, Sawasdee N, Guizouarn H, Pellissier B, Borgese F, et al. Band 3 Edmonton I, a novel mutant of the anion exchanger 1 causing spherocytosis and distal renal tubular acidosis. *Biochem J.* 2010;426:379–88, <http://dx.doi.org/10.1042/BJ20091525>.
 38. Kaitwatcharakchai C, Vasuvattakul S, Yenchitsomanus PT, Thuwajit P, Malasit P, Chuawatana D, et al. Distal renal tubular acidosis and high urine carbon dioxide tension in a patient with southeast Asian ovalocytosis. *Am J Kidney Dis.* 1999;33:1147–52, [http://dx.doi.org/10.1016/s0272-6386\(99\)70154-x](http://dx.doi.org/10.1016/s0272-6386(99)70154-x).
 39. Vasuvattakul S. Molecular approach for distal renal tubular acidosis associated AE1 mutations. *Electrolyte Blood Press.* 2010;8:25–31, <http://dx.doi.org/10.5049/EBP.2010.8.1.25>.