

The role of Fibroblast Growth Factor 23 in chronic kidney disease-mineral and bone disorder

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Nefrologia 2013;33(6):835-44

doi10.3265/Nefrologia.pre2013.Jul.12091

ABSTRACT

Fibroblast Growth Factor 23 (FGF-23) is a bone-derived hormone involved in the regulation of phosphate homeostasis. FGF-23 levels are extremely elevated in Chronic Kidney Disease (CKD) and there is evidence supporting the role of this hormone in the pathogenesis of Chronic Kidney Disease-Mineral and Bone Disorder (CKD-MBD). Furthermore, recent data associates FGF-23 with the pathogenesis of systemic complications of CKD-MBD. The increasing evidence that the consequences of abnormal mineral metabolism are not restricted to bone disease changed the approach to the pathophysiology and treatment of disturbed bone and mineral metabolism in CKD patients. FGF-23 has been proposed to be the initial adaptive response in early CKD to protect the organism from the adverse effects of phosphate retention. Increased levels of FGF-23 observed in CKD patients are associated with cardiovascular mortality risk and was shown to mediate direct, "off-target" toxicity to the heart. This report aims to review the relevant aspects of the physiology of FGF-23 in bone biology and mineral homeostasis and the role of FGF-23 in the pathophysiology of CKD-MBD and its clinical implications.

Keywords: Chronic kidney disease (CKD). FGF-23. Mineral and bone metabolism. Bone metabolism. CKD-MBD.

INTRODUCTION

Chronic Kidney Disease (CKD) is a worldwide public health issue associated with several adverse outcomes, including cardiovascular disease and premature death.¹ Although CKD

El papel del factor de crecimiento de fibroblastos 23 en el trastorno mineral y óseo asociado a la enfermedad renal crónica

RESUMEN

El factor de crecimiento fibroblástico 23 (FGF-23) es una hormona derivada del hueso que participa en la regulación de la homeostasis del fósforo. Los niveles de FGF-23 se encuentran extremadamente elevados en la enfermedad renal crónica y existe evidencia del papel de esta hormona en la patogénesis de los trastornos óseos y minerales en esta situación. Más aún, datos recientes implican al FGF-23 en la patogénesis de otras complicaciones sistémicas asociadas a las alteraciones óseo-minerales de la enfermedad renal crónica. La evidencia creciente de que las alteraciones del metabolismo mineral no se limitan a la enfermedad ósea ha acentuado el interés por la patofisiología y el tratamiento de las alteraciones del metabolismo mineral en la enfermedad renal crónica. Se ha propuesto que el aumento de FGF-23 es la respuesta inicial en los estadios precoces de la enfermedad renal crónica a la necesidad de proteger al organismo de los efectos adversos de la retención de fósforo. Estos aumentos de FGF-23 se asocian al riesgo creciente de mortalidad cardiovascular en los enfermos renales crónicos y son mediadores directos de toxicidad cardíaca. En esta revisión procuramos presentar aspectos relevantes de la fisiología del FGF-23 en la biología ósea y en la homeostasis mineral, así como en la fisiopatología de la enfermedad renal crónica y sus implicaciones clínicas.

Palabras clave: Enfermedad renal crónica (ERC). FGF-23. Metabolismo óseo-mineral. Metabolismo óseo. TMO-ERC.

encompasses a broad spectrum of different pathophysiologic processes, disorders of bone and mineral metabolism develop in virtually all patients as their kidney function declines.² The discovery of Fibroblast Growth Factor 23 (FGF-23) happened in the sequence of the study of several rare hypophosphatemic disorders like *X-linked Hypophosphatemia* or *Tumor Induced Osteomalacia*.³ The elevated levels of FGF-23 observed in CKD patients^{4,5} were noticed very soon after the identification of this hormone. FGF-23 is a phosphaturic hormone increasing renal phosphate excretion and also

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reduces 1,25-dihydroxyvitamin D synthesis at the kidney level and increases its degradation. In early CKD, FGF-23 may be the first marker of disrupted mineral metabolism⁶ and the pivotal player in the pathophysiology and progression of Chronic Kidney Disease - Mineral and Bone Disorder (CKD-MBD), which is associated not only with bone abnormalities but also cardiovascular disease and vascular calcification.⁵ Recently, FGF-23 has been associated with several adverse clinical outcomes, such as the development of secondary hyperparathyroidism⁶ and left ventricular hypertrophy (LVH),⁷ as well as faster deterioration of renal function⁸ and increased mortality.⁹

CHRONIC KIDNEY DISEASE-MINERAL AND BONE DISORDER (CKD-MBD): A PARADIGM SHIFT

In the past, the disruption of mineral homeostasis in CKD was seen as the alteration of calcium and phosphorus levels and the dysregulation of the PTH and 1,25-dihydroxyvitamin D axis, which would eventually lead to secondary hyperparathyroidism¹⁰ and bone disease. Together, these abnormalities were known as “renal osteodystrophy”.¹¹ With the increasing evidence that the deleterious consequences of disturbed mineral metabolism in CKD were not restricted to the development of parathyroid and bone disease but were also associated to vascular calcification and cardiovascular morbidity and mortality,^{5,12} the Kidney Disease Improving Global Outcomes working group gathered in 2006 to rework the definition of renal osteodystrophy. Together, these abnormalities in calcium, phosphorus, PTH and vitamin D metabolism, bone turnover, mineralization, volume, strength and linear growth and cardiovascular or soft tissue calcification, all due to CKD, were re-named “Chronic Kidney Disease - Mineral and Bone Disorder”, reflecting their systemic and complex nature.⁵ This new concept also fits well in terms of pathophysiology, since the mechanisms behind the various manifestations are closely entwined.¹¹

OVERVIEW OF FGF-23

FGF-23 is a 32 kDa phosphate-regulating hormone secreted mainly by osteocytes and osteoblasts.¹³ This hormone inhibits renal phosphate reabsorption and reduces systemic levels of 1,25-dihydroxyvitamin D,¹⁴ leading to diminished phosphate load as a consequence of decrease renal tubular reabsorption and absorption from the gastrointestinal tract. FGF-23 performs these hypophosphatemic actions through the downregulation of type II sodium-phosphate cotransporters (NaPhT-2a and NaPhT-2c) localized in the apical membrane of the proximal convoluted tubule (PCT),^{14,15} inhibition of 25-hydroxyvitamin D 1 α -hydroxylase and the stimulation of 25-hydroxyvitamin D-24-hydroxylase.^{14,16} Also FGF-23 is able to interact with PTH. FGF-23 directly suppresses PTH mRNA in vitro and decreases serum PTH in vivo.¹⁷ FGF-23 is

elevated in several hereditary and acquired hypophosphatemic disorders, including *X-linked hypophosphatemia*, *Autosomal Dominant Hypophosphatemic Rickets (ADHR)* or *Tumor Induced Osteomalacia (TIO)*. These disorders share a common phenotype of phosphaturia, hypophosphatemia and rickets/osteomalacia.¹⁸ In contrast, *Hyperphosphatemic Familial Tumoral Calcinosis* is characterized by soft tissue calcifications, hyperphosphatemia and elevated 1,25-dihydroxyvitamin D levels due to reduced levels of active FGF-23.¹⁹

FGF-23 has affinity to various receptors such as Fibroblast Growth Factor Receptor 1C, 3C and 4. The regulation of phosphate reabsorption is mediated by its binding to the Fibroblast Growth Factor Receptor 1C (FGFR-1C) and to the co-receptor α -Klotho, which increases the binding affinity of FGF-23 for FGFRs. FGFR 3C and 4 seem to play a greater role in the regulation of 1,25-dihydroxyvitamin D metabolism.²⁰

The co-receptor α -Klotho is a 130 kDa transmembrane protein expressed mainly in the distal convoluted tube (DCT), parathyroid and vascular tissue.²¹ Another report does not support Klotho-mediated FGF-23 effects in the vasculature²² but confirmative studies in humans are needed. Klotho was accidentally discovered as a mutated gene in mice with a premature aging disorder²³ and its name comes from the Greek mythology: Klotho was the goddess who spun the thread of life. Conveniently, its anti-aging properties were afterward demonstrated in animal studies.²⁴ Further investigation demonstrated that when α -Klotho is absent, the activity of FGF-23 is essentially suppressed.²⁵ In addition to membrane-bound α -Klotho, the co-receptor also exists in two circulating forms, generated by different processes - alternate RNA splicing or proteolytic cleavage.²⁶ Circulating α -Klotho is also capable of enabling FGF-23's signal transduction, but at a lower level than the membrane-bound form.¹⁸

FGF-23 diminishes α -Klotho expression,²⁷ maybe through reductions in 1,25-dihydroxyvitamin D levels, which is known to stimulate α -Klotho expression.²⁸ The proteolytic cleavage product of α -Klotho was implicated in the direct regulation of FGF-23.²⁹ This form of Klotho is capable of inducing FGF-23 production in osteocytes in vivo, which suggest the presence of a new endocrine feedback loop in mineral metabolism.³⁰ In CKD, FGF-23 is markedly elevated¹⁵ and α -Klotho is severely depressed.³¹ Several other actions of Klotho not related to FGF-23 have been described³² but those are outside of the scope of this review.

FGF-23 AND BONE BIOLOGY

There is increasing evidence linking bone biology and FGF-23. This hormone appears to coordinate renal phosphate handling to match bone mineralization and remodeling,^{13,33} which influences both the influx and efflux of calcium and

phosphate from bone.³⁴ Also, many studies suggest that FGF-23 may have a direct effect on bone with a direct inhibition of osteoblast differentiation and matrix mineralization *in vitro*,³⁵ while a complete lack of FGF-23 results in impaired skeletal mineralization even with adequate phosphate, calcium and 1,25-dihydroxyvitamin D levels in mature animals.³⁶ Consistent with these findings is the disruption of the Wnt-signaling pathway, which is responsible for osteoblast proliferation and bone matrix mineralization, noted in mice with excess FGF-23 expression in bone.³⁷ In FGF-23 null mice the administration of a phosphate-deficient diet reversed hyperphosphatemia completely and greatly improved survival but the bone mineralization defects persisted.³⁸ In a double FGF-23 and NaPhT-2a knock-out mouse model, the skeletal phenotype is similar to FGF-23 null mice, although the phosphate levels are not elevated.³⁹ These studies not only suggest that the bone defects related to the absence of FGF-23 are independent of its actions on phosphate homeostasis, but also suggest that FGF-23 has a physiologic role in bone biology.

The finding of numerous single gene mutations leading to alterations in bone remodeling and increased FGF-23,³⁷ also favor this hypothesis. Examples of these mutations are those affecting PHEX (*phosphate-regulating gene with homologies to endoptidases on the X-chromosome*)⁴⁰ and members of the SIBLING protein family, such as DMP1 (*dentin matrix acidic phosphoprotein 1*)⁴¹ and MEPE (*matrix extracellular phosphoglycoprotein*).³³ The exact mechanism by which these proteins regulate FGF-23 is yet to be determined. PHEX and DMP1 seem to exert its inhibitory effect through the FGFR1 receptor in the osteocyte.⁴² Increases in MEPE³³ and loss of PHEX or DMP1 expression⁴² are both associated to defects in skeletal mineralization and increased levels of FGF-23. FGF-23 activity is also regulated at the protein level in which glycosylation by GALNT3 (*GalNAc Transferase 3*) protects it from the proteolytic cleavage by Furin,¹⁹ allowing the secretion of intact FGF-23 (iFGF-23) into circulation.⁴³ iFGF-23 is biologically active, while the product of cleavage by Furin, the C-terminal form (cFGF-23) is not,³ but may be able to block the bioactivity of the intact form.⁴⁴

Despite no detectable Klotho mRNA on bone,²³ very recently the expression of FGF-23 and α -Klotho mRNA and protein were detected in porcine growth plate chondrocytes,⁴⁵ which further highlights a potential role for FGF-23 in bone physiology. Shalhoub and colleagues demonstrated that supraphysiologic levels of FGF-23, in the presence of soluble α -Klotho, increase osteoblastic *MC3T3.E1* cell proliferation and inhibit bone mineralization *in vitro*²⁶ but, contrary to what was described by Wang et al.,³⁵ over-expression of FGF-23 alone did not affect proliferation or mineralization.²⁶ Samadfam et al., showed in mouse models that a sustained reduction in bone turnover increased FGF-23,⁴⁶ which is concordant with its effect on bone mineralization. In addition, Wesseling-Perry and colleagues observed an inverse relationship between FGF-23 and osteoid thickness, osteoid

maturation time and mineralization lag time in pediatric stage 5 CKD patients with secondary hyperparathyroidism, suggesting an association between FGF-23 and improved indices of skeletal mineralization in this population.⁴⁷ Although these data seems contradictory it clearly shows the complexity of the mechanisms by which FGF-23 is associated with skeletal mineralization. Further research is required to clearly define FGF-23's role on bone biology and its possible implications in CKD-MBD.

SYSTEMIC REGULATION OF FGF-23: A COMPLEX SCENARIO

FGF-23 is regulated by phosphorus,^{48,49} 1,25-dihydroxyvitamin D⁵⁰ and probably also by PTH¹⁷ (Figure 1). A dietary phosphorus increase and 1,25-dihydroxyvitamin D augment FGF-23 levels, while dietary restriction of phosphorus has the opposite effect.⁴⁸⁻⁵⁰ The observation of the effect of phosphorus on FGF-23 is mostly derived from feeding studies in healthy humans and evidence of a direct effect is currently lacking. In fact, when serum phosphate was elevated through non-dietary sources, FGF-23 stayed unaltered,⁵¹ and in the feeding studies the changes in serum phosphate did not precede changes in FGF-23, as would be expected.^{48,49} This phosphate-sensing mechanism that regulates FGF-23 remains a mystery and might involve an unidentified phosphate sensor¹⁸ or be the indirect result of alterations in bone mineralization due to phosphate.⁵² The effects of 1,25-dihydroxyvitamin D on FGF-23 are thought to be controlled by a vitamin D response element (VDRE) in the FGF-23 promoter gene.⁵³ Although this VDRE is present in both mouse⁵³ and rat,⁵⁴ and there is support for a gatekeeper effect,⁵⁵ no analogous region has yet been found within the FGF-23 promoter of the human gene.¹⁸ The complexity of this system is also supported by the finding that in the absence of the Vitamin D Receptor, the increase in dietary phosphate does not increase FGF-23.⁵⁶

Like 1,25-dihydroxyvitamin D, PTH seems to share a negative endocrine feedback loop with FGF-23. PTH stimulates the synthesis of FGF-23 both directly⁵⁷ and indirectly via PTH-mediated increases in 1,25-dihydroxyvitamin D⁵⁸ while PTH secretion is inhibited via a FGFR-Klotho-dependent pathway.¹⁷ However FGF-23 does not prevent the development of hyperparathyroidism (HPT) and there is a strong link between the FGF-23 increase and the severity of HPT in CKD,²⁵ suggesting a possible causality effect.²³ Furthermore, in the absence of PTH, FGF-23 levels are high rather than diminished,⁵⁹ suggesting that PTH is not essential to increase FGF-23 secretion in humans. Also, in hypoparathyroidism FGF-23 action seems to be crippled, perhaps because of the absence of PTH or the low levels of 1,25-dihydroxyvitamin D.⁵⁹ Indeed, inducing hypoparathyroidism with cinacalcet in patients with a FGF-23 excess disease like *TIO* attenuates FGF-23 action⁶⁰ and cinacalcet seems to lower FGF-23 levels in CKD.⁶¹ This dynamic interplay between PTH and FGF-23 needs to be further

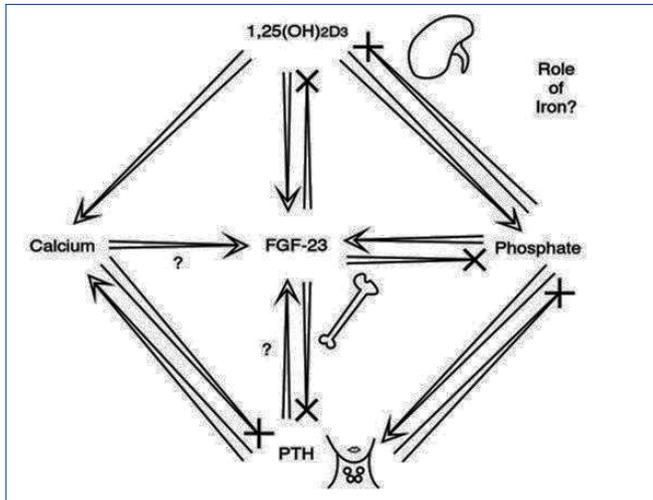


Figure 1. The complexity of the Kidney-Parathyroid-Bone Axis that regulates FGF-23.

FGF-23 is directly regulated by phosphate, 1,25-dihydroxyvitamin D, PTH and possibly by calcium. A role for Iron is emerging. See the text for more details. Legend: Arrowed line (→): directly increases serum levels. Crossed line (→x): directly decreases serum levels. ?: Requires further studies to fully enlighten this interaction.

clarified but it is already well established that both hormones are needed to maintain serum phosphate homeostasis.

A high calcium diet increased FGF-23 secretion both in wild type mice and in vitamin D receptor-ablated mice.⁵⁶ This supports a stimulatory effect of calcium on FGF-23 secretion since other possible stimuli (PTH and 1,25-dihydroxyvitamin D) were decreased because of hypercalcemia or blunted in vitamin D receptor-ablated mice. In addition, studies show that non-calcium based phosphate binders like sevelamer, reduce FGF-23 more than calcium-based phosphate binders despite comparable reductions on serum phosphate,⁶² supporting the hypothesis of a direct effect of calcium on FGF-23. Finally, in the hypercalcemia of malignancy, FGF-23 levels are elevated despite PTH being suppressed.⁶³ Hypocalcemia was associated with low FGF-23 in rats fed with a low calcium and low vitamin D diet,⁶⁴ despite high serum levels of PTH and 1,25-dihydroxyvitamin D.⁶⁵ This effect was reversed by the administration of calcium. Although we wait for further elucidation, this stimulatory effect of calcium on FGF-23 is undoubtedly important and should be taken into account in the therapeutic management of CKD-MBD in the years to come.

Finally, a potential role for iron in the regulation of FGF-23 is emerging. In the CKD population, especially in patients undergoing dialysis, parental iron therapy with erythropoietin is widely used to manage anemia. After some studies showing an association between iron therapy and bone mineral abnormalities, investigators began to speculate that intravenous iron administration could be responsible for an increase in FGF-23 levels and play a

role in the development of hypophosphatemic osteomalacia during iron therapy.^{66,67} In fact, intravenous iron is capable of increasing FGF-23 secretion and induce hypophosphatemia in non-CKD patients.⁶⁸ In stage 5 CKD population results are contradictory. Takeda *et al.*, concluded that intravenous saccharated ferric oxide induces a small increase in the already elevated FGF-23 levels in hemodialysis patients⁶⁹ while Deger *et al.*, unexpectedly found a negative relationship between iron administration and serum iFGF-23 levels in a dialysis population.⁷⁰ There is also contradictory data suggesting that iron deficiency increases FGF-23 expression in both wild-type mice and healthy individuals.^{71,72} In these populations, the serum iron levels negatively correlated with cFGF-23 but not with iFGF-23 while in ADHR subjects low serum iron was associated to an increase in both cFGF-23 and iFGF-23, which results in hypophosphatemia.^{71,72} Since ADHR is caused by an FGF-23 mutation that renders it resistant to cleavage,⁷³ it can be postulated that iron deficiency does not increase iFGF-23 in healthy subjects because it is cleaved by Furin into smaller fragments which are then released and able to be detected with the C-terminal assay.⁷⁴ Better understanding of the bone cell pathways related to this iron/FGF-23 interaction is required to determine the actual role of iron and the real importance of FGF-23 processing in the regulation of active FGF-23 levels.

PATHOPHYSIOLOGY OF CKD-MBD: IS IT FGF-23-CENTRIC?

There is still a lot of controversy regarding the pathophysiology of CKD-MBD. Actually there is no consensus on what induces/exacerbates CKD-MBD or the “right” order of appearance of the various factors involved. Several studies showed that FGF-23 levels are elevated in patients with various degrees of CKD and rise progressively as renal function declines^{4,75} (Figure 2).⁷⁶ The exact CKD stage where FGF-23 levels start to increase is not completely clear and different studies have some inconsistent results.^{4,74} Also, elevated levels of FGF-23 were consistently associated with higher serum phosphate, higher fractional excretion of phosphate, lower estimated glomerular filtration rate (eGFR) and lower levels of 1,25-dihydroxyvitamin D.^{4,74} This latter association is independent of eGFR. It has been proposed that the rise of FGF-23 in early CKD is due to a compensatory response to intermittent increases in enteral phosphate burden because of the impaired renal excretion,¹⁰ diminished expression of α -Klotho that induces resistance to FGF-23¹ or due to alterations in osteocyte biology^{77,78} that somehow stimulates FGF-23 secretion directly.⁴⁶ Others suggest that PTH levels increase first and result in elevated FGF-23 concentrations.^{17,48}

The current data shows that FGF-23 expression in osteocytes is increased in early CKD, occurring as early as stage 2 at a time when 1,25-dihydroxyvitamin D, PTH, calcium and phosphate

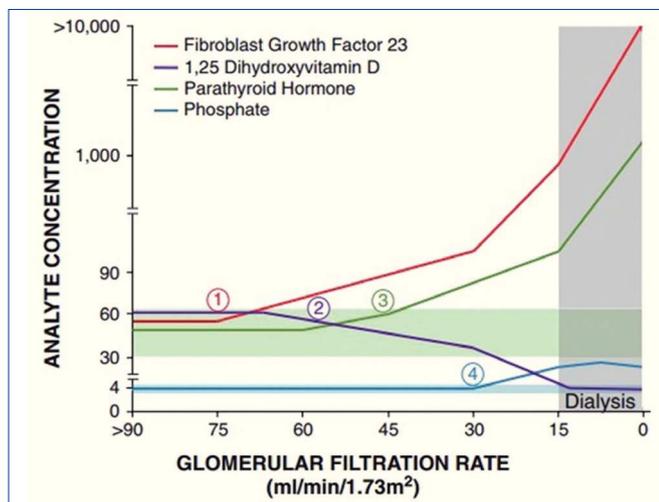


Figure 2. Fibroblast Growth Factor 23 (1), 1,25-dihydroxyvitamin D (2), Parathyroid Hormone (3) and Phosphate (4) levels (y axis) according to Glomerular Filtration Rate (x axis).

The y axis represents circulating levels of the individual analytes with the temporal changes in and normal ranges of individual analytes color coded (FGF-23 (RU/ml) in red, 1,25-dihydroxyvitamin D (pg/ml) in purple, PTH (pg/ml) in green and phosphate (mg/dl) in blue). The increase in FGF23 is the earliest alteration in mineral metabolism in CKD (1). Gradually increasing FGF23 levels cause the early decline in 1,25-dihydroxyvitamin D levels (2) that free PTH from feedback inhibition (3). Phosphate levels remain unaltered until late stages of CKD (4). See Figure 3 and the text for further details. This figure was adapted and reproduced with permission from the Journal of the American Society of Nephrology.

are still within normal range.⁷⁹ DMP1 is also up-regulated at this stage,⁷⁷ which appears to be an effort of the osteocyte to suppress FGF-23 production. In addition, serum FGF-23 levels are augmented in 30-40% of adults with an estimated GFR of 70-90ml/min/1.73m² and serum phosphate was actually a bit lower than in subjects with higher eGFR.⁶ This reduction in serum phosphate combined with increased urinary fractional excretion of phosphate, suggest they are product of FGF-23 elevation.⁶ Since reduced α -Klotho expression is expected to cripple the excretion of phosphate,²³ the lack of an increase in serum phosphate in early CKD also does not favor the FGF-23 resistance model of CKD-MBD. Also, PTH concentrations rose in only 10% of this same population.⁶

Although the initial trigger to the elevation of FGF-23 in early CKD is yet to be demonstrated, there is evidence to support that the early elevation in FGF-23 is the initial adaptive response in CKD⁸⁰ and the first measurable biochemical evidence of disturbed mineral ion homeostasis in CKD-MBD.¹⁰ We can hypothesize (Figure 3) that this early FGF-23 elevation leads to a decrease in 1,25-dihydroxyvitamin D levels, as a consequence of inhibition of 25-hydroxyvitamin D 1 α -hydroxylase rather than the loss of renal mass, as was

traditionally thought.⁷⁵ This decrease in 1,25-dihydroxyvitamin D levels is responsible for a secondary increase in PTH due to impaired intestinal calcium absorption (the transient hypocalcemia stimulates the calcium-sensing receptors in the parathyroid gland (PTG)) and loss of 1,25-dihydroxyvitamin D's negative feedback on the PTG. PTH would maintain normocalcemia at the expense of stimulating bone remodeling, the reduction of calcium excretion by the kidney and by increasing 25-hydroxyvitamin D 1 α -hydroxylase activity. This augment in bone remodeling, not only increases bone calcium but also bone phosphate efflux, which has to be excreted by a declining number of functional nephrons. This increase in phosphate can further stimulate FGF-23 secretion. PTH is also capable of directly stimulating FGF-23 production.⁵⁷ Furthermore, PTH-mediated effects on bone remodeling may lead to local alterations in bone biology that further amplify FGF-23 expression.⁵² This additional FGF-23 facilitates phosphate wasting by the kidney, but when

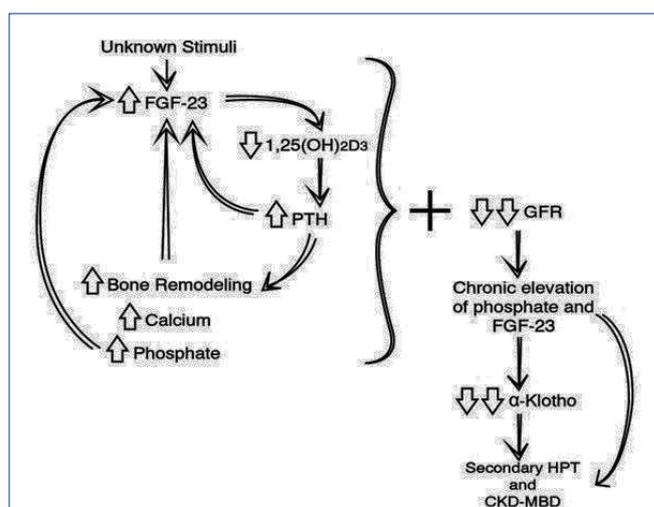


Figure 3. Current hypothesis about the pathophysiology of Chronic Kidney Disease-Mineral and Bone Disorder. Unknown stimuli cause an early elevation in FGF-23 serum levels which leads to a decrease in 1,25-dihydroxyvitamin D levels, as a consequence of inhibition of 25-hydroxyvitamin D 1 α -hydroxylase. This causes a secondary increase in PTH through various mechanisms. PTH maintains normocalcemia mainly at the expense of stimulating bone remodeling and the reduction of calcium excretion by the kidney. This augment in bone remodeling not only increases serum calcium but also bone phosphate efflux, which can stimulate FGF-23 secretion. PTH is capable of directly increasing FGF-23 levels. Bone remodeling may further amplify FGF-23 expression. When GFR becomes severely decreased, phosphate continuously stays above normal levels, leading to a positive feedback loop, further increasing FGF-23 and PTH levels. This chronic elevation of phosphate and FGF-23 leads to down-regulation of α -Klotho expression. The combination of the chronic elevation of FGF-23 and phosphate and reduced α -Klotho expression leads to Secondary Hyperparathyroidism and CKD-MBD. See the text for further details.

renal function becomes severely affected, phosphate chronically stays above normal levels, further suppressing 25-hydroxyvitamin D 1 α -hydroxylase activity and increasing PTH secretion (the elevation in intracellular phosphorus contributes to the increase in lifespan of PTH mRNAs¹¹). In parallel, this state of chronic FGF-23 excess induces a down-regulation in α -Klotho expression, which clearly has a critical role in the progression of CKD-MBD.¹ This reduced α -Klotho expression along with reduced FGFR expression in the PTG⁸¹ is responsible for the resistance to the inhibition of PTH expression by FGF-23 and contributes to the coexistence of secondary increases in both FGF-23 and PTH in CKD-MBD.⁸² In addition, the expression of the receptors for calcium⁸³ and 1,25-dihydroxyvitamin D⁸⁴ also decline, eventually leading to the development of an PTG secreting PTH autonomously of the suppressing stimuli.¹¹ Thus, in the late stages of CKD, low levels of 1,25-dihydroxyvitamin D, hypocalcemia, hyperphosphatemia, reduced expression of α -Klotho and high levels of FGF-23 all contribute to the development of secondary hyperparathyroidism⁸⁵ and CKD-MBD.

FGF-23 AND CLINICAL OUTCOMES

Besides the deleterious effects of the disruption of mineral metabolism, FGF-23 is an independent predictor of mortality,⁹ progression of renal disease,⁸ LVH⁷ and possibly vascular dysfunction⁸⁶ in CKD patients. Faul et al. recently demonstrated supraphysiologic levels of FGF-23 similar to those seen in late CKD were able to induce left ventricular hypertrophy in an α -Klotho knockout mice.⁸⁷ Although, we still await other studies to confirm this effect, this results support the hypothesis of α -Klotho-independent and direct “off-target” toxicity of FGF-23. It may also explain the consistent association between FGF-23 and LVH and establish a bridge between higher FGF-23 and greater mortality risk in the CKD population. In fact, components of the uremic cardiomyopathy,⁸⁸ such as progressive ventricular failure, arrhythmia and sudden death (in which LVH is an important mechanism), are more common causes of death in advanced CKD than acute myocardial infarction.⁸⁹ Even if there is no consistently robust association between FGF-23 and artery calcification,¹ α -Klotho deficiency has been implicated in the pathophysiology of vascular calcification,⁹⁰ which may be an indirect effect of chronically elevated FGF-23 levels in CKD.

CLINICAL IMPLICATIONS IN CKD-MBD

The hypothesis presented on the chapter *Pathophysiology of CKD-MBD: is it FGF-23-centric?* has obvious limitations. It does not explain why FGF-23 increases in the first place and how FGF-23 and PTH interact in the early stages of CKD. It does not take into account the possible effects of hypocalcemia or the role of iron on FGF-23 expression. Furthermore, it offers a “static” view of the bone mineral metabolism and presupposes

that every patient with CKD progresses at the same speed. Yet, if it proves true, this integrated concept of the pathophysiology of CKD-MBD will have clinical implications. This hypothesis suggests that early CKD does not represent a true 1,25-dihydroxyvitamin D deficient state, but an adaptive response to protect against hyperphosphatemia, which means that FGF-23 may be a biomarker for earlier interventions in the CKD population. This could become an interesting tool to help interpreting low 1,25-dihydroxyvitamin D levels in the elderly population (e.g. a decrease due to FGF-23 in subclinical CKD versus true deficiency).⁸⁰ Also, at advanced stages of CKD the main stimulus for FGF23 elevation may be bone remodeling, supporting other interventional strategies targeting bone rather than the administration of 1,25-dihydroxyvitamin D.

Because 1,25-dihydroxyvitamin D further stimulates FGF23 production, 1,25-dihydroxyvitamin D sparing therapeutic strategies could be important, such as combined low dose of active vitamin D compounds and calcimimetics, which are proven to lower FGF-23 in stage 5 CKD patients.⁹¹ Still, there is a paradox to be solved since epidemiological data has shown a survival advantage for CKD patients treated with active vitamin D analogs,^{92,93} and at the same time, vitamin D also increases FGF-23 and phosphate serum levels, contributing to calcifications,⁹⁴ left ventricular hypertrophy⁷ and mortality.⁹ Wolf, M. proposed that the variability of FGF-23 response to active vitamin D therapy between populations may be the key to unravel this potential contradiction.¹ Well designed, prospective, randomized, controlled clinical trials are clearly necessary comparing the effect of different therapeutic protocols in hard outcomes such as mortality.

CONCLUSION

In little more than a decade, the discovery of FGF-23 reformed our knowledge about mineral homeostasis in both health and disease. Still, we are just beginning to unravel the complex pathophysiologic process of CKD-MBD and many questions remain to be answered. What is the real role of FGF-23 in bone biology? What is the precipitating factor for the early FGF-23 increase in CKD? What is the role of iron? Does FGF-23 directly induce LVH? Is the current standard of care optimal? Our hope is that the resolution of these and other matters will help designing randomized clinical trials which will end in meaningful improvements in clinical outcomes for CKD patients in the future.

Conflicts of interest

Receive honoraria as speaker: honorarios por ponencias: João M. Frazão: Amgen, Sanofi, Abbott.

KEY CONCEPTS

1. FGF-23 inhibits renal phosphate reabsorption and reduces the levels of 1,25-dihydroxyvitamin D.
2. FGF-23 might have a physiologic role in bone biology and is complexly regulated both locally and systemically.
3. FGF-23 is the initial adaptive response in early CKD and plays an important role in the pathogenesis of the systemic complications of CKD-MBD.
4. The deleterious effects of the FGF-23 excess in CKD are due to both its physiologic actions and "off-target" toxicity.

REFERENCES

1. Wolf M. Update on fibroblast growth factor 23 in chronic kidney disease. *Kidney Int* 2012;82(7):737-47.
2. Fukagawa M, Akizawa T. Calcium abnormalities of dialysis patients. *J Bone Miner Metab* 2006;24(2):160.
3. Shimada T, Muto T, Urakawa I, Yoneya T, Yamazaki Y, Okawa K, et al. Mutant FGF-23 responsible for autosomal dominant hypophosphatemic rickets is resistant to proteolytic cleavage and causes hypophosphatemia in vivo. *Endocrinology* 2002;143(8):3179-82.
4. Larsson T, Nisbeth U, Ljunggren O, Juppner H, Jonsson KB. Circulating concentration of FGF-23 increases as renal function declines in patients with chronic kidney disease, but does not change in response to variation in phosphate intake in healthy volunteers. *Kidney Int* 2003;64(6):2272-9.
5. Moe S, Drueke T, Cunningham J, Goodman W, Martin K, Olgaard K, et al. Definition, evaluation, and classification of renal osteodystrophy: a position statement from Kidney Disease: Improving Global Outcomes (KDIGO). *Kidney Int* 2006;69(11):1945-53.
6. Isakova T, Wahl P, Vargas GS, Gutierrez OM, Scialla J, Xie H, et al. Fibroblast growth factor 23 is elevated before parathyroid hormone and phosphate in chronic kidney disease. *Kidney Int* 2011;79(12):1370-8.
7. Canziani ME, Tomiyama C, Higa A, Draibe SA, Carvalho AB. Fibroblast growth factor 23 in chronic kidney disease: bridging the gap between bone mineral metabolism and left ventricular hypertrophy. *Blood Purif* 2011;31(1-3):26-32.
8. Fliser D, Kollerits B, Neyer U, Ankerst DP, Lhotta K, Lingenhel A, et al. Fibroblast growth factor 23 (FGF23) predicts progression of chronic kidney disease: the Mild to Moderate Kidney Disease (MMKD) Study. *J Am Soc Nephrol* 2007;18(9):2600-8.
9. Isakova T, Xie H, Yang W, Xie D, Anderson AH, Scialla J, et al. Fibroblast growth factor 23 and risks of mortality and end-stage renal disease in patients with chronic kidney disease. *JAMA* 2011;305(23):2432-9.
10. Wesseling-Perry K, Juppner H. The osteocyte in CKD: New concepts regarding the role of FGF23 in mineral metabolism and systemic complications. *Bone* 2012;54(2):222-9.
11. Mac Way F, Lessard M, Lafage-Proust MH. Pathophysiology of chronic kidney disease-mineral and bone disorder. *Joint Bone Spine* 2012;79(6):544-9.
12. Bargman J, Skorecki K. In: Longo D, Fauci, A (eds.). *Harrison's Principles of Internal Medicine*, Chap. 280. 18th Edition ed. United States of America: Mc-Graw Hill; 2012.
13. Liu S, Zhou J, Tang W, Jiang X, Rowe DW, Quarles LD. Pathogenic role of Fgf23 in Hyp mice. *Am J Physiol Endocrinol Metab* 2006;291(1):E38-49.
14. Shimada T, Hasegawa H, Yamazaki Y, Muto T, Hino R, Takeuchi Y, et al. FGF-23 is a potent regulator of vitamin D metabolism and phosphate homeostasis. *J Bone Miner Res* 2004;19(3):429-35.
15. Cozzolino M, Mazzaferro S. The fibroblast growth factor 23: a new player in the field of cardiovascular, bone and renal disease. *Curr Vasc Pharmacol* 2010;8(3):404-11.
16. Saito H, Kusano K, Kinosaki M, Ito H, Hirata M, Segawa H, et al. Human fibroblast growth factor-23 mutants suppress Na⁻-dependent phosphate co-transport activity and 1 α ,25-dihydroxyvitamin D₃ production. *J Biol Chem* 2003;278(4):2206-11.
17. Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, et al. The parathyroid is a target organ for FGF23 in rats. *J Clin Invest* 2007;117(12):4003-8.
18. Bhattacharyya N, Chong WH, Gafni RI, Collins MT. Fibroblast growth factor 23: state of the field and future directions. *Trends Endocrinol Metab* 2012;23(12):610-8.
19. Topaz O, Shurman DL, Bergman R, Indelman M, Ratajczak P, Mizrachi M, et al. Mutations in GALNT3, encoding a protein involved in O-linked glycosylation, cause familial tumoral calcinosis. *Nat Genet* 2004;36(6):579-81.
20. Li H, Martin A, David V, Quarles LD. Compound deletion of Fgfr3 and Fgfr4 partially rescues the Hyp mouse phenotype. *Am J Physiol Endocrinol Metab* 2011;300(3):E508-17.
21. Donate-Correa J, Mora-Fernández C, Martínez-Sanz R, Muros-de-Fuentes M, Pérez H, Meneses-Perez B, et al. Expression of FGF23/KLOTHO system in human vascular tissue. *Int J Cardiol* 2013;165(1):179-83.
22. Lindberg K, Olauson H, Amin R, Ponnusamy A, Goetz R, Taylor RF, et al. Arterial klotho expression and FGF23 effects on vascular calcification and function. *PLoS One* 2013;8(4):e60658.
23. Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, et al. Mutation of the mouse klotho gene leads to a syndrome resembling ageing. *Nature* 1997;390(6655):45-51.

24. Kurosu H, Yamamoto M, Clark JD, Pastor JV, Nandi A, Gurnani P, et al. Suppression of aging in mice by the hormone Klotho. *Science* 2005;309(5742):1829-33.
25. Urakawa I, Yamazaki Y, Shimada T, Iijima K, Hasegawa H, Okawa K, et al. Klotho converts canonical FGF receptor into a specific receptor for FGF23. *Nature* 2006;444(7120):770-4.
26. Shalhoub V, Ward SC, Sun B, Stevens J, Renshaw L, Hawkins N, et al. Fibroblast growth factor 23 (FGF23) and alpha-klotho stimulate osteoblastic MC3T3.E1 cell proliferation and inhibit mineralization. *Calcif Tissue Int* 2011;89(2):140-50.
27. Marsell R, Krajsnik T, Goransson H, Ohlsson C, Ljunggren O, Larsson TE, et al. Gene expression analysis of kidneys from transgenic mice expressing fibroblast growth factor-23. *Nephrol Dial Transplant* 2008;23(3):827-33.
28. Tsujikawa H, Kurotaki Y, Fujimori T, Fukuda K, Nabeshima Y. Klotho, a gene related to a syndrome resembling human premature aging, functions in a negative regulatory circuit of vitamin D endocrine system. *Mol Endocrinol* 2003;17(12):2393-403.
29. Smith RC, O'Bryan LM, Farrow EG, Summers LJ, Clinkenbeard EL, Roberts JL, et al. Circulating alphaKlotho influences phosphate handling by controlling FGF23 production. *J Clin Invest* 2012;122(12):4710-5.
30. Juppner H, Wolf M. alphaKlotho: FGF23 coreceptor and FGF23-regulating hormone. *J Clin Invest* 2012;122(12):4336-9.
31. Koh N, Fujimori T, Nishiguchi S, Tamori A, Shiomi S, Nakatani T, et al. Severely reduced production of klotho in human chronic renal failure kidney. *Biochem Biophys Res Commun* 2001;280(4):1015-20.
32. Donate-Correa J, Muros-de-Fuentes M, Mora-Fernández C, Navarro-González JF. FGF23/Klotho axis: phosphorus, mineral metabolism and beyond. *Cytokine Growth Factor Rev* 2012;23(1-2):37-46.
33. Liu S, Rowe PS, Vierthaler L, Zhou J, Quarles LD. Phosphorylated acidic serine-aspartate-rich MEPE-associated motif peptide from matrix extracellular phosphoglycoprotein inhibits phosphate regulating gene with homologies to endopeptidases on the X-chromosome enzyme activity. *J Endocrinol* 2007;192(1):261-7.
34. Sato T, Tominaga Y, Ueki T, Goto N, Matsuoka S, Katayama A, et al. Total parathyroidectomy reduces elevated circulating fibroblast growth factor 23 in advanced secondary hyperparathyroidism. *Am J Kidney Dis* 2004;44(3):481-7.
35. Wang H, Yoshiko Y, Yamamoto R, Minamizaki T, Kozai K, Tanne K, et al. Overexpression of fibroblast growth factor 23 suppresses osteoblast differentiation and matrix mineralization in vitro. *J Bone Miner Res* 2008;23(6):939-48.
36. Sitara D, Razzaque MS, Hesse M, Yoganathan S, Taguchi T, Erben RG, et al. Homozygous ablation of fibroblast growth factor-23 results in hyperphosphatemia and impaired skeletogenesis, and reverses hypophosphatemia in Phex-deficient mice. *Matrix Biol* 2004;23(7):421-32.
37. Liu S, Tang W, Fang J, Ren J, Li H, Xiao Z, et al. Novel regulators of Fgf23 expression and mineralization in Hyp bone. *Mol Endocrinol* 2009;23(9):1505-18.
38. Stubbs JR, Liu S, Tang W, Zhou J, Wang Y, Yao X, et al. Role of hyperphosphatemia and 1,25-dihydroxyvitamin D in vascular calcification and mortality in fibroblastic growth factor 23 null mice. *J Am Soc Nephrol* 2007;18(7):2116-24.
39. Sitara D, Kim S, Razzaque MS, Bergwitz C, Taguchi T, Schuler C, et al. Genetic evidence of serum phosphate-independent functions of FGF-23 on bone. *PLoS Genet* 2008;4(8):e1000154.
40. Liu S, Guo R, Simpson LG, Xiao ZS, Burnham CE, Quarles LD. Regulation of fibroblastic growth factor 23 expression but not degradation by PHEX. *J Biol Chem* 2003;278(39):37419-26.
41. Feng JQ, Ward LM, Liu S, Lu Y, Xie Y, Yuan B, et al. Loss of DMP1 causes rickets and osteomalacia and identifies a role for osteocytes in mineral metabolism. *Nat Genet* 2006;38(11):1310-5.
42. Martin A, Liu S, David V, Li H, Karydis A, Feng JQ, et al. Bone proteins PHEX and DMP1 regulate fibroblastic growth factor Fgf23 expression in osteocytes through a common pathway involving FGF receptor (FGFR) signaling. *FASEB J* 2011;25(8):2551-62.
43. Ichikawa S, Baujat G, Seyahi A, Garoufali AG, Imel EA, Padgett LR, et al. Clinical variability of familial tumoral calcinosis caused by novel GALNT3 mutations. *Am J Med Genet A* 2010;152A(4):896-903.
44. Goetz R, Nakada Y, Hu MC, Kurosu H, Wang L, Nakatani T, et al. Isolated C-terminal tail of FGF23 alleviates hypophosphatemia by inhibiting FGF23-FGFR-Klotho complex formation. *Proc Natl Acad Sci U S A* 2010;107(1):407-12.
45. Raimann A, Ertl DA, Helmreich M, Sagmeister S, Egerbacher M, Haeusler G. Fibroblast growth factor 23 and Klotho are present in the growth plate. *Connect Tissue Res* 2013;54(2):108-17.
46. Samadfam R, Richard C, Nguyen-Yamamoto L, Bolivar I, Goltzman D. Bone formation regulates circulating concentrations of fibroblast growth factor 23. *Endocrinology* 2009;150(11):4835-45.
47. Wesseling-Perry K, Pereira RC, Wang H, Elashoff RM, Sahney S, Gales B, et al. Relationship between plasma fibroblast growth factor-23 concentration and bone mineralization in children with renal failure on peritoneal dialysis. *J Clin Endocrinol Metab* 2009;94(2):511-7.
48. Antonucci DM, Yamashita T, Portale AA. Dietary phosphorus regulates serum fibroblast growth factor-23 concentrations in healthy men. *J Clin Endocrinol Metab* 2006;91(8):3144-9.
49. Gutierrez OM, Wolf M, Taylor EN. Fibroblast growth factor 23, cardiovascular disease risk factors, and phosphorus intake in the health professionals follow-up study. *Clin J Am Soc Nephrol* 2011;6(12):2871-8.
50. Wesseling-Perry K, Pereira RC, Sahney S, Gales B, Wang HJ, Elashoff R, et al. Calcitriol and doxercalciferol are equivalent in controlling bone turnover, suppressing parathyroid hormone, and increasing fibroblast growth factor-23 in secondary hyperparathyroidism. *Kidney Int* 2011;79(1):112-9.
51. Ito N, Fukumoto S, Takeuchi Y, Takeda S, Suzuki H, Yamashita T, et al. Effect of acute changes of serum phosphate on fibroblast growth factor (FGF)23 levels in humans. *J Bone Miner Metab* 2007;25(6):419-22.
52. Mirams M, Robinson BG, Mason RS, Nelson AE. Bone as a source of FGF23: regulation by phosphate? *Bone* 2004;35(5):1192-9.
53. Liu S, Tang W, Zhou J, Stubbs JR, Luo Q, Pi M, et al. Fibroblast growth factor 23 is a counter-regulatory phosphaturic hormone for vitamin D. *J Am Soc Nephrol* 2006;17(5):1305-15.
54. Barthel TK, Mathern DR, Whitfield GK, Haussler CA, Hopper HA, Hsieh JC, et al. 1,25-Dihydroxyvitamin D3/VDR-mediated induction of FGF23 as well as transcriptional control of other bone anabolic and catabolic genes that orchestrate the regulation of phosphate and calcium mineral metabolism. *J Steroid Biochem Mol Biol* 2007;103(3-5):381-8.

55. Ohnishi M, Nakatani T, Lanske B, Razzaque MS. Reversal of mineral ion homeostasis and soft-tissue calcification of klotho knockout mice by deletion of vitamin D 1alpha-hydroxylase. *Kidney Int* 2009;75(11):1166-72.
56. Shimada T, Yamazaki Y, Takahashi M, Hasegawa H, Urakawa I, Oshima T, et al. Vitamin D receptor-independent FGF23 actions in regulating phosphate and vitamin D metabolism. *Am J Physiol Renal Physiol* 2005;289(5):F1088-95.
57. Lavi-Moshayoff V, Wasserman G, Meir T, Silver J, Naveh-Many T. PTH increases FGF23 gene expression and mediates the high-FGF23 levels of experimental kidney failure: a bone parathyroid feedback loop. *Am J Physiol Renal Physiol* 2010;299(4):F882-9.
58. López I, Rodríguez-Ortiz ME, Almadén Y, Guerrero F, de Oca AM, Pineda C, et al. Direct and indirect effects of parathyroid hormone on circulating levels of fibroblast growth factor 23 in vivo. *Kidney Int* 2011;80(5):475-82.
59. Gupta A, Winer K, Econs MJ, Marx SJ, Collins MT. FGF-23 is elevated by chronic hyperphosphatemia. *J Clin Endocrinol Metab* 2004;89(9):4489-92.
60. Geller JL, Khosravi A, Kelly MH, Riminucci M, Adams JS, Collins MT. Cinacalcet in the management of tumor-induced osteomalacia. *J Bone Miner Res* 2007;22(6):931-7.
61. Finch JL, Tokumoto M, Nakamura H, Yao W, Shahnazari M, Lane N, et al. Effect of paricalcitol and cinacalcet on serum phosphate, FGF-23, and bone in rats with chronic kidney disease. *Am J Physiol Renal Physiol* 2010;298(6):F1315-22.
62. Yilmaz MI, Sonmez A, Saglam M, Yaman H, Kilic S, Eyiletten T, et al. Comparison of calcium acetate and sevelamer on vascular function and fibroblast growth factor 23 in CKD patients: a randomized clinical trial. *Am J Kidney Dis* 2012;59(2):177-85.
63. Singh RJ, Kumar R. Fibroblast growth factor 23 concentrations in humoral hypercalcemia of malignancy and hyperparathyroidism. *Mayo Clin Proc* 2003;78(7):826-9.
64. Rodríguez-Ortiz ME, Lopez I, Munoz-Castaneda JR, Martinez-Moreno JM, Ramirez AP, Pineda C, et al. Calcium deficiency reduces circulating levels of FGF23. *J Am Soc Nephrol* 2012;23(7):1190-7.
65. Rodríguez M, López I, Muñoz J, Aguilera-Tejero E, Almadén Y. FGF23 and mineral metabolism, implications in CKD-MBD. *Nefrologia* 2012;32(3):275-8.
66. Schouten BJ, Hunt PJ, Livesey JH, Frampton CM, Soule SG. FGF23 elevation and hypophosphatemia after intravenous iron polymaltose: a prospective study. *J Clin Endocrinol Metab* 2009;94(7):2332-7.
67. Schouten BJ, Doogue MP, Soule SG, Hunt PJ. Iron polymaltose-induced FGF23 elevation complicated by hypophosphatemic osteomalacia. *Ann Clin Biochem* 2009;46(Pt 2):167-9.
68. Shimizu Y, Tada Y, Yamauchi M, Okamoto T, Suzuki H, Ito N, et al. Hypophosphatemia induced by intravenous administration of saccharated ferric oxide: another form of FGF23-related hypophosphatemia. *Bone* 2009;45(4):814-6.
69. Takeda Y, Komaba H, Goto S, Fujii H, Umezu M, Hasegawa H, et al. Effect of intravenous saccharated ferric oxide on serum FGF23 and mineral metabolism in hemodialysis patients. *Am J Nephrol* 2011;33(5):421-6.
70. Deger SM, Erten Y, Pasaoglu OT, Derici UB, Reis KA, Onec K, et al. The effects of iron on FGF23-mediated Ca-P metabolism in CKD patients. *Clin Exp Nephrol* 2013;17(3):416-23.
71. Imel EA, Peacock M, Gray AK, Padgett LR, Hui SL, Econs MJ. Iron modifies plasma FGF23 differently in autosomal dominant hypophosphatemic rickets and healthy humans. *J Clin Endocrinol Metab* 2011;96(11):3541-9.
72. Farrow EG, Yu X, Summers LJ, Davis SI, Fleet JC, Allen MR, et al. Iron deficiency drives an autosomal dominant hypophosphatemic rickets (ADHR) phenotype in fibroblast growth factor-23 (Fgf23) knock-in mice. *Proc Natl Acad Sci U S A* 2011;108(46):E1146-55.
73. ADHR Consortium. Autosomal dominant hypophosphatemic rickets is associated with mutations in FGF23. *Nat Genet* 2000;26(3):345-8.
74. Bhattacharyya N, Wiench M, Dumitrescu C, Connolly BM, Bugge TH, Patel HV, et al. Mechanism of FGF23 processing in fibrous dysplasia. *J Bone Miner Res* 2012;27(5):1132-41.
75. Gutierrez O, Isakova T, Rhee E, Shah A, Holmes J, Collierone G, et al. Fibroblast growth factor-23 mitigates hyperphosphatemia but accentuates calcitriol deficiency in chronic kidney disease. *J Am Soc Nephrol* 2005;16(7):2205-15.
76. Wolf M. Forging forward with 10 burning questions on FGF23 in kidney disease. *J Am Soc Nephrol* 2010;21(9):1427-35.
77. Pereira RC, Juppner H, Azucena-Serrano CE, Yadin O, Salusky IB, Wesseling-Perry K. Patterns of FGF-23, DMP1, and MEPE expression in patients with chronic kidney disease. *Bone* 2009;45(6):1161-8.
78. Sabbagh Y, Gracioli FG, O'Brien S, Tang W, dos Reis LM, Ryan S, et al. Repression of osteocyte Wnt/beta-catenin signaling is an early event in the progression of renal osteodystrophy. *J Bone Miner Res* 2012;27(8):1757-72.
79. Levin A, Bakris GL, Molitch M, Smulders M, Tian J, Williams LA, et al. Prevalence of abnormal serum vitamin D, PTH, calcium, and phosphorus in patients with chronic kidney disease: results of the study to evaluate early kidney disease. *Kidney Int* 2007;71(1):31-8.
80. Quarles LD. Role of FGF23 in vitamin D and phosphate metabolism: implications in chronic kidney disease. *Exp Cell Res* 2012;318(9):1040-8.
81. Komaba H, Goto S, Fujii H, Hamada Y, Kobayashi A, Shibuya K, et al. Depressed expression of Klotho and FGF receptor 1 in hyperplastic parathyroid glands from uremic patients. *Kidney Int* 2010;77(3):232-8.
82. Galitzer H, Ben-Dov IZ, Silver J, Naveh-Many T. Parathyroid cell resistance to fibroblast growth factor 23 in secondary hyperparathyroidism of chronic kidney disease. *Kidney Int* 2010;77(3):211-8.
83. Gogusev J, Duchambon P, Hory B, Giovannini M, Goureau Y, Sarfati E, et al. Depressed expression of calcium receptor in parathyroid gland tissue of patients with hyperparathyroidism. *Kidney Int* 1997;51(1):328-36.
84. Brown AJ, Dusso A, Lopez-Hilker S, Lewis-Finch J, Grooms P, Slatopolsky E. 1,25-(OH)2D receptors are decreased in parathyroid glands from chronically uremic dogs. *Kidney Int* 1989;35(1):19-23.
85. Gutierrez OM, Mannstadt M, Isakova T, Rauh-Hain JA, Tamez H, Shah A, et al. Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. *N Engl J Med* 2008;359(6):584-92.
86. Mirza MA, Larsson A, Lind L, Larsson TE. Circulating fibroblast growth factor-23 is associated with vascular dysfunction in the community. *Atherosclerosis* 2009;205(2):385-90.
87. Faul C, Amaral AP, Oskouei B, Hu MC, Sloan A, Isakova T, et al. FGF23 induces left ventricular hypertrophy. *J Clin Invest* 2011;121(11):4393-408.

88. Gross ML, Ritz E. Hypertrophy and fibrosis in the cardiomyopathy of uremia--beyond coronary heart disease. *Semin Dial* 2008;21(4):308-18.
89. Shastri S, Tangri N, Tighiouart H, Beck GJ, Vlagopoulos P, Ornt D, et al. Predictors of sudden cardiac death: a competing risk approach in the hemodialysis study. *Clin J Am Soc Nephrol* 2012;7(1):123-30.
90. Hu MC, Shi M, Zhang J, Quinones H, Griffith C, Kuro-o M, et al. Klotho deficiency causes vascular calcification in chronic kidney disease. *J Am Soc Nephrol* 2011;22(1):124-36.
91. Wetmore JB, Liu S, Krebill R, Menard R, Quarles LD. Effects of cinacalcet and concurrent low-dose vitamin D on FGF23 levels in ESRD. *Clin J Am Soc Nephrol* 2010;5(1):110-6.
92. Kovesdy CP, Ahmadzadeh S, Anderson JE, Kalantar-Zadeh K. Association of activated vitamin D treatment and mortality in chronic kidney disease. *Arch Intern Med* 2008;168(4):397-403.
93. Shoben AB, Rudser KD, de Boer IH, Young B, Kestenbaum B. Association of oral calcitriol with improved survival in nondialyzed CKD. *J Am Soc Nephrol* 2008;19(8):1613-9.
94. O'Neill WC, Lomashvili KA. Recent progress in the treatment of vascular calcification. *Kidney Int* 2010;78(12):1232-9.